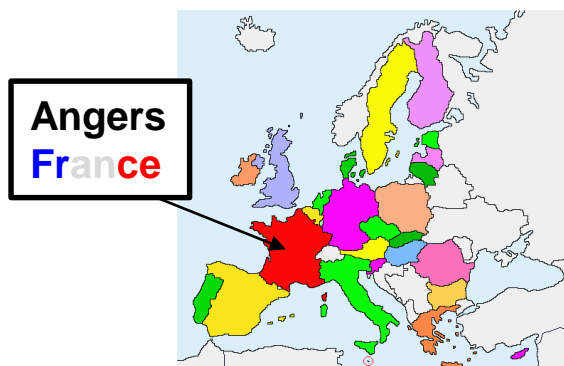


# Carbon modified sensors dedicated to organic micropollutants analysis and innovant waters treatment (WT) aid



**M. PONTI , professor  
Angers University**

<http://geihp.univ-angers.fr>

KEYNOTE SPEAKER, 27<sup>th</sup> Novembre 9h45

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## General outlines

**1/ GENERAL INTRODUCTION**

**2/ COMMERCIALIZED ELECTROCHEMICAL SENSORS**

**3/ THE PROBLEM OF ORGANIC MICROPOLLUTANTS IN WATER RESSOURCES**

**4/ CASE STUDIES 1 : Electrochemical sensors development**

⇒ **UME, CPE, GCE, PGE DEVELOPMENTS DEDICATED TO ORGANIC MICROPOLLUTANTS**

**5/ CASE STUDIES 2 : Electrochemical analysis and WWT processes**

⇒ **BIOREACTORS DEVELOPMENT + NANOFILTRATION**

⇒ **ANTIBIOFILMS STRATEGIES ON UME & CPE**

# 1/ GENERAL INTRODUCTION

need to measure, detect, monitor

# MEDICAL / HEALTH

Self-diagnosis (glucose, lactate)



Multi-analysis for emergencies



for the patient in his hospital room



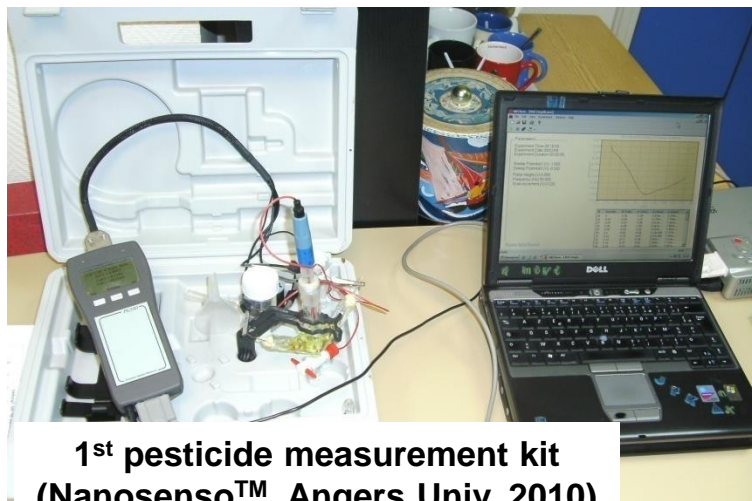
Multi-analysis for biological laboratories

Studies interactions drug receptor

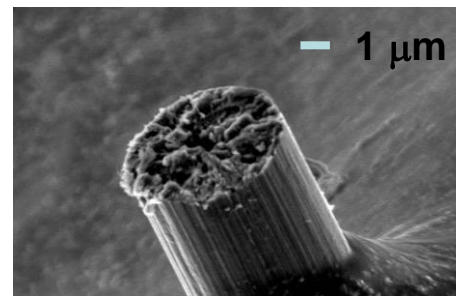


# ENVIRONMENTAL ANALYSIS

**OUTSIDE  
THE  
LAB**



**1<sup>st</sup> pesticide measurement kit  
(Nanosenso™, Angers Univ. 2010)**



**UME**

Nanosenso™, 2010, GA&P/GEPEA

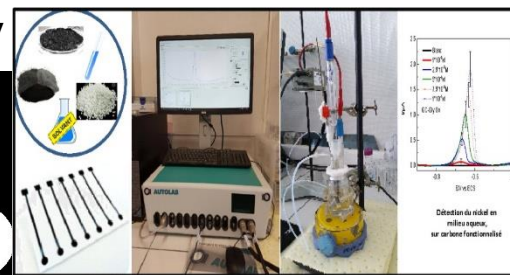


**INSIDE  
THE  
LAB**

**Bacterial biosensor, BOD  
measurement**

**(Biosensors) GEPEA 2008 (Nantes)**

Orléans University



**SPE**



# Analysis for what ?

## DIAGNOSIS

not only...

**WATERS TREATMENT PROCESSES AID  
FOR THEIR INTENSIFICATION**

## **2/ ELECTROCHEMICAL SENSORS**

### **POTENTIOMETRIC, AMPEROMETRIC**

# Commercialized Sensors (1)

Kind of electrode	Species
<p style="text-align: center;">Solid Membrane</p> <ul style="list-style-type: none"> <li>❖ glass</li> <li>❖ monocrystal</li> <li>❖ compacted solids (powder)</li> </ul>	<p><math>H^+</math>, <math>Na^+</math>, <math>K^+</math>, (<math>Li^+</math>, <math>Rb^+</math>, <math>Ag^+</math>)</p> <p><math>F^-</math></p> <p><math>Ag^+</math>, <math>Cd^{2+}</math>, <math>Cu^{2+}</math>, <math>Pb^{2+}</math>, <math>S^{2-}</math>, <math>Cl^-</math>, <math>Br^-</math>, <math>I^-</math>, <math>CN^-</math>, <math>SCN^-</math></p>
<p style="text-align: center;">Liquid membrane</p> <ul style="list-style-type: none"> <li>❖ neutral transport</li> <li>❖ ionics exchange</li> </ul>	<p><math>K^+</math>, <math>NH_4^+</math>, <math>Na^+</math>, (<math>Li^+</math>, <math>Ca^{2+}</math>)</p> <p><math>Ca^{2+}</math>, <math>Cu^{2+}</math>, <math>Cl^-</math>, <math>BF_4^-</math>, <math>Mg^{2+}</math>, <math>NO_3^-</math>, <math>ClO_4^-</math>,</p> <p>Organics anions and cations</p>
<p style="text-align: center;">Gaz sensors</p>	<p><math>NH_3</math>, <math>CO_2</math>, <math>SO_2</math>, <math>HCN</math>, <math>H_2S</math></p>

Limit of detection (LOD) :  $10^{-6}$  mol/L



# Commercialized amperometric Sensors (2)

Kind of sensors	Companies
<p style="text-align: center;">Solid membrane</p> <ul style="list-style-type: none"> <li>❖ NO (oxydation in NO<sup>+</sup>)</li> <li>❖ O<sub>2</sub> (reduction in H<sub>2</sub>O)</li> <li>❖ inorganics micropollutants (reduction of Pb<sup>2+</sup>; Cd<sup>2+</sup>; Hg<sup>2+</sup>)</li> <li>❖ <b>Organics micropollutants →</b></li> </ul>	<p>WPI (US)</p> <p>HACH LANGE (US)</p> <p>Some commercialized devices</p> <ul style="list-style-type: none"> <li>• Autolab (<a href="http://www.ecochemie.nl">http://www.ecochemie.nl</a>)</li> <li>• Palmsens (<a href="http://www.palmsens.com">http://www.palmsens.com</a>)</li> <li>• Orignalys (<a href="http://www.origalys.com/fr/">http://www.origalys.com/fr/</a>)</li> <li>• Metrohm (<a href="http://www.metrohm.com">http://www.metrohm.com</a>)</li> </ul> <p><b>no devices commercialized</b></p>
<p style="text-align: center;">Gaz detectors</p>	<p>O<sub>2</sub>, H<sub>2</sub>S</p>

Limit of detection : 10<sup>-12</sup> mol/L

### 3/ THE PROBLEM OF ORGANIC MICROPOLLUTANTS IN WATER RESSOURCES

All **chemical substances** detected in the form of **traces** and which are of **human origin**. There are 3 categories of micropollutants: organic, inorganic and organometallic [1].

Most of those compounds comes from industrial syntheses. These are products with a wide variety of uses : pesticides, biocides, cleaning agents ...

**Emerging micropollutants are the drug molecules and their metabolites.**

#### **A PUBLIC AND ANALYTICAL HEALTH ISSUE**

They are scattered in **all compartments of the environment**

They are found in trace amounts (**submicromolar concentrations**)

#### **A NEED**

**Monitoring** aquatic environments, our food and our body

#### **A NECESSITY**

**Analysis and treatment of contaminated compartments**

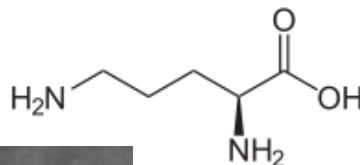
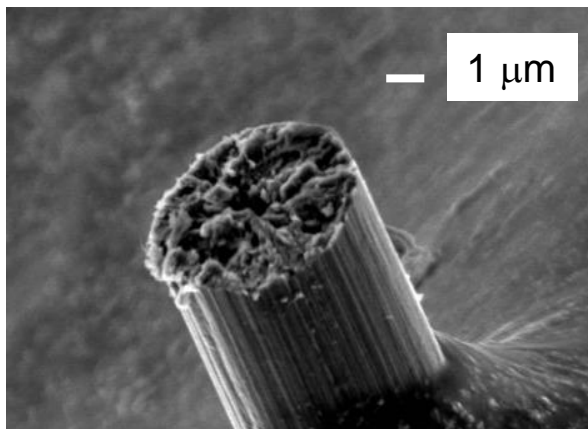
[1] **Chèvre N., Erkman S.**, Alerte aux micropolluants, Pesticides, biocides, détergents, médicaments et autres substances chimiques dans l'environnement, Nature et Environnement, **Collection Le Savoir Suisse**, Presses polytechniques et universitaires romandes, **2011**, 143 p., ISBN 9-78-2880-749385

**4/ CASE STUDIES 1 : Electrochemical sensors development**  
**UME AND CPE SENSORS DEDICATED TO ORGANIC**  
**MICROPOLLUTANTS**

# Analyzes of Molecules of **Biological** and **Environmental** Interests in my team (Hospital center Angers)

**NO**

Ultra-microelectrode (**UME**)  
with C fiber



**Ornithine**

PAP

DCF

HQ

MPT

PNP

phenol

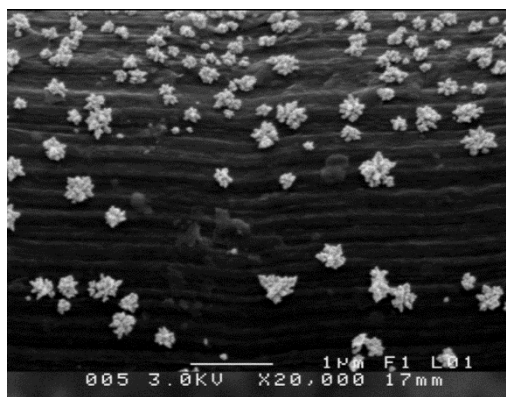
4-HBz

APAP



**MNP**

**Fenitrothion**



Gold nanosensors (AuNPs)  
on C UME

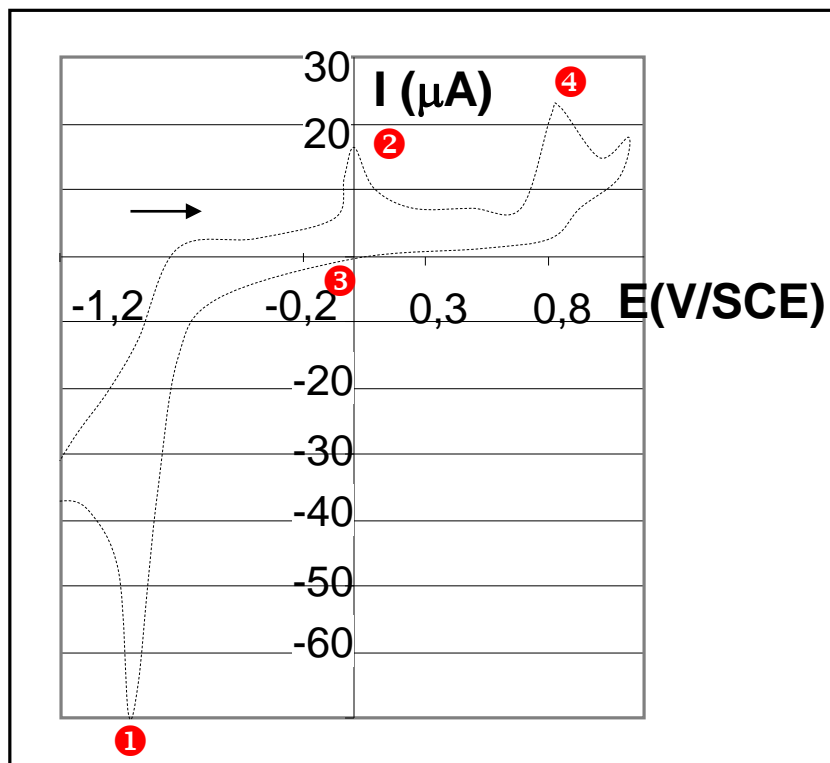
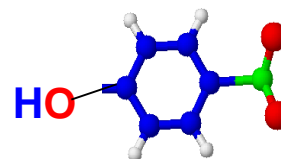


**PGE**  
(pencil  
graphite  
electrode)

**glucose**



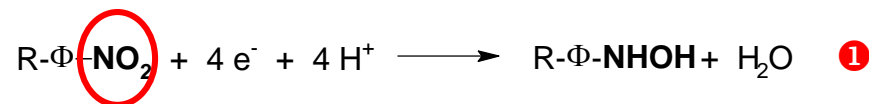
# Electroactivity of PNP on GCE



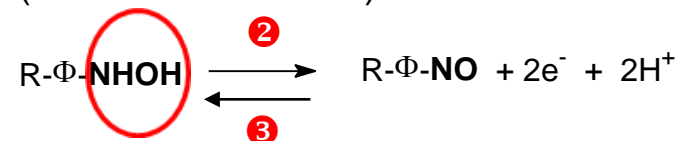
## Cyclic Voltammetry of PNP

- GCE
- acetate buffer (5.2, 0.1 M)
- potential scan rate : 100 mV.s<sup>-1</sup>
- [PNP] = 20 mg.L<sup>-1</sup>

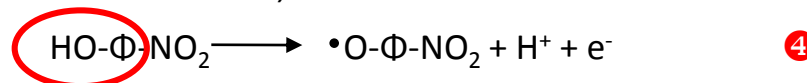
Irreversible reduction of -NO<sub>2</sub>  
(**direct detection**):



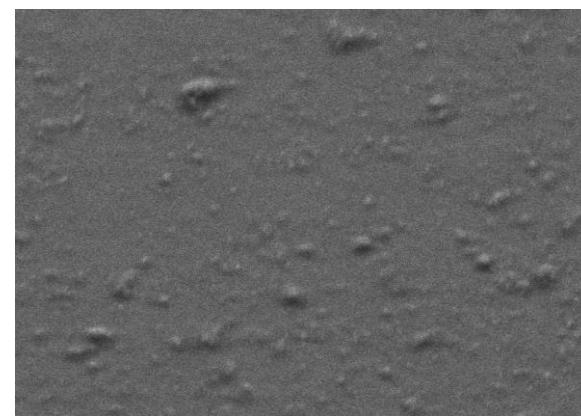
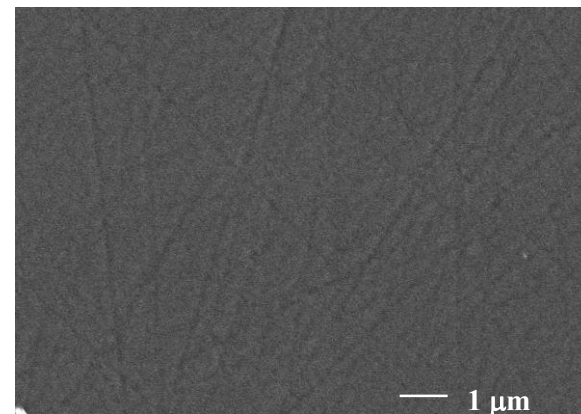
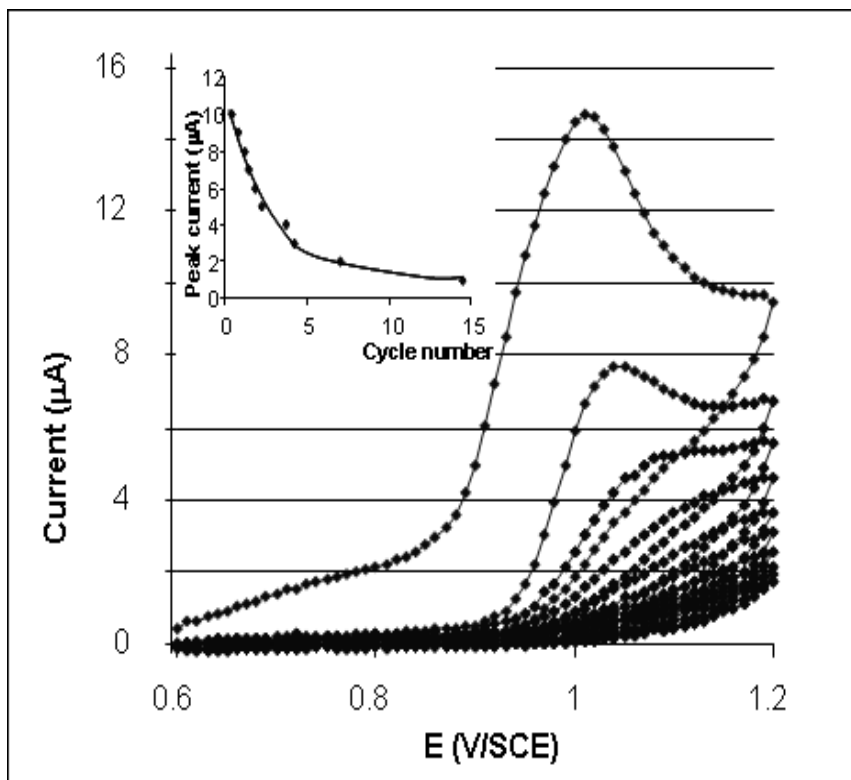
Reversible oxidation of -NHOH  
(**indirect detection**):



Irreversible oxidation of -Φ-OH  
(**direct detection**):



# Electrode **passivation** with PNP oxidation

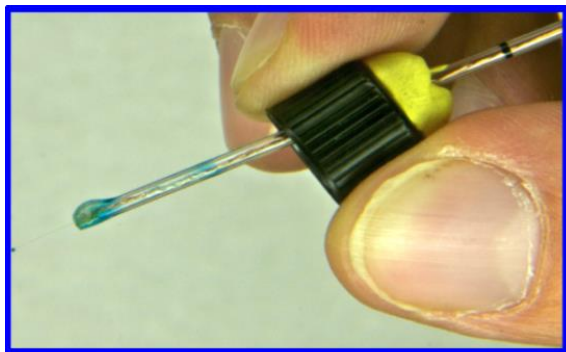


BE CAREFULL WITH THE **FORMATION**  
**OF POLYPHENOL FILM !**

- électrolyte : ACETATE BUFFER (pH=5;2 ; 0;1M)
- SCAN RATE : 100 mV.s<sup>-1</sup>
- [PNP] = 100 mg.L<sup>-1</sup> ( 5.10<sup>-4</sup> mol.L<sup>-1</sup>)

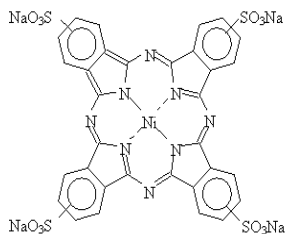
# Improving the performances of UMEs' : PNP case

- 1- UME elaboration + quality control



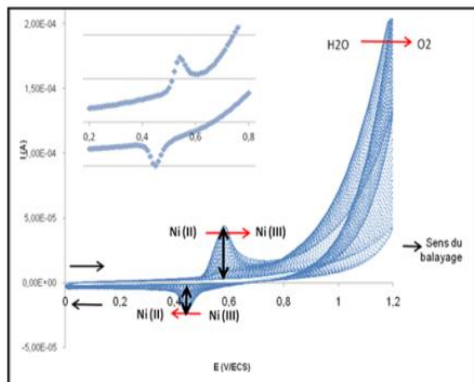
« *Homemade* » UME

- 2- UME modification by a film of phthalocyanine (p-NiTSPc)

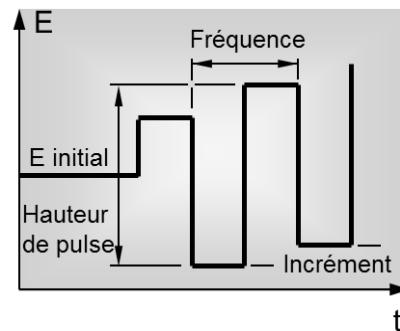


NiTSPc

Alcaline electrodeposition pH = 12



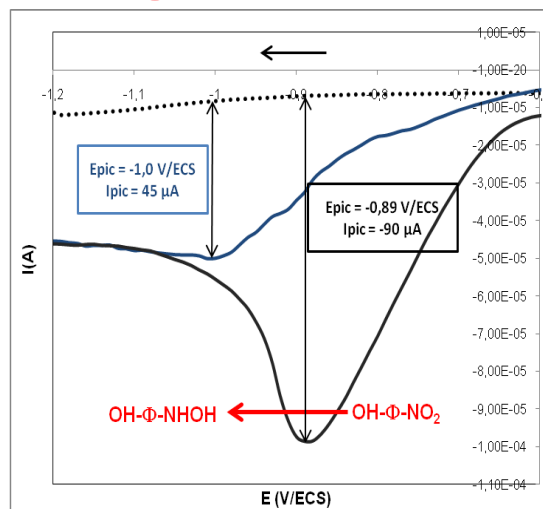
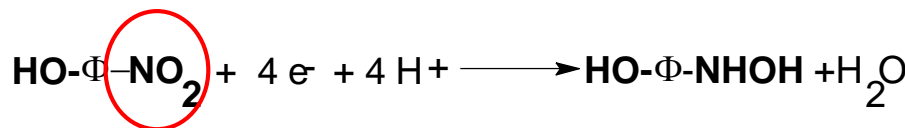
- 3- Square wave voltammetry



E initial : -0.2 V/ECS  
 Frequency : 60 Hz,  
 Pulse height: 90 mV  
 Potential incre.: 20 mV

Decreased the capacitive current

- 4- PNP analysis



--- baseline  
 — no p-NiTSPc  
 — With p-NiTSPc

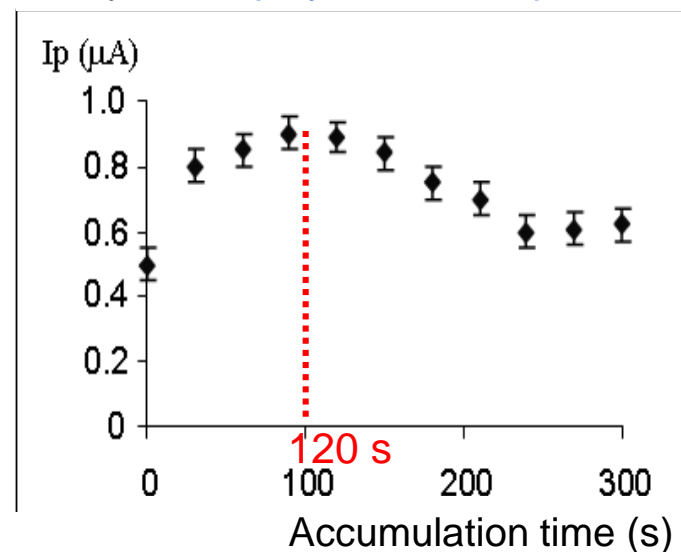
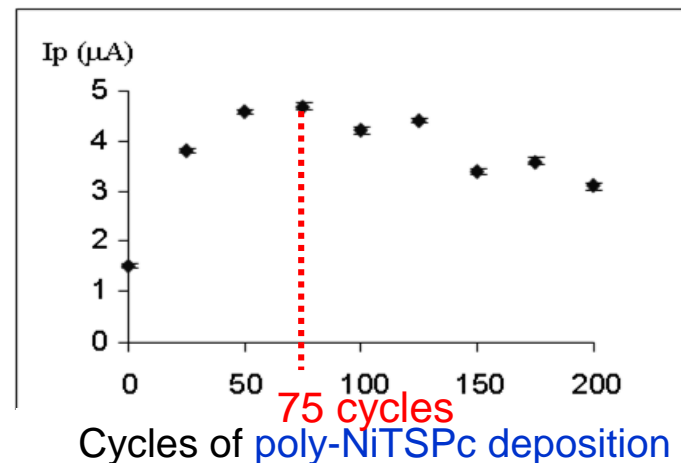
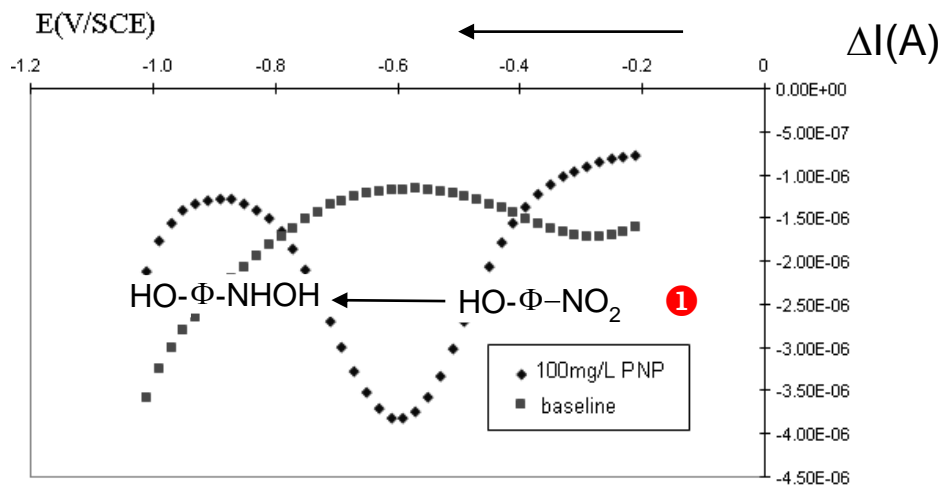
Electrocatalytic effect

# 5-Higher sensitivity



UME :  
C/p-NiTSPc

E initial : -0.2 V/ECS  
Frequency : 60 Hz  
Pulse height : 90 mV  
Pot. Increm. : 20 mV



Operating conditions	LOD ( μg/L ± 0,05)
C cleaned	100.00
C/poly-NiTSPc (75) No accum. time	25.00
C/poly-NiTSPc (75) 2 min. of accum.	<b>0.10</b>

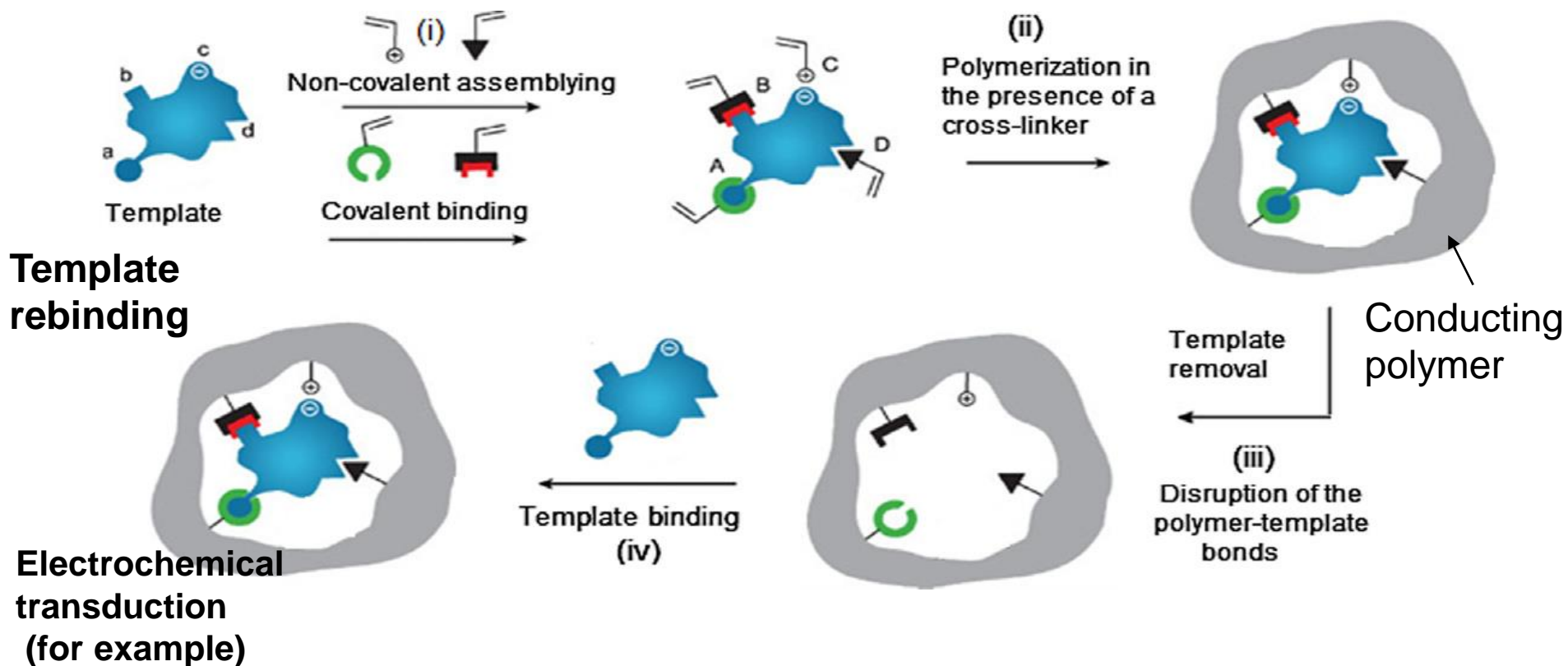


## 6- Analyzes comparison between GC / MS and modified UME of estuarine and surface waters

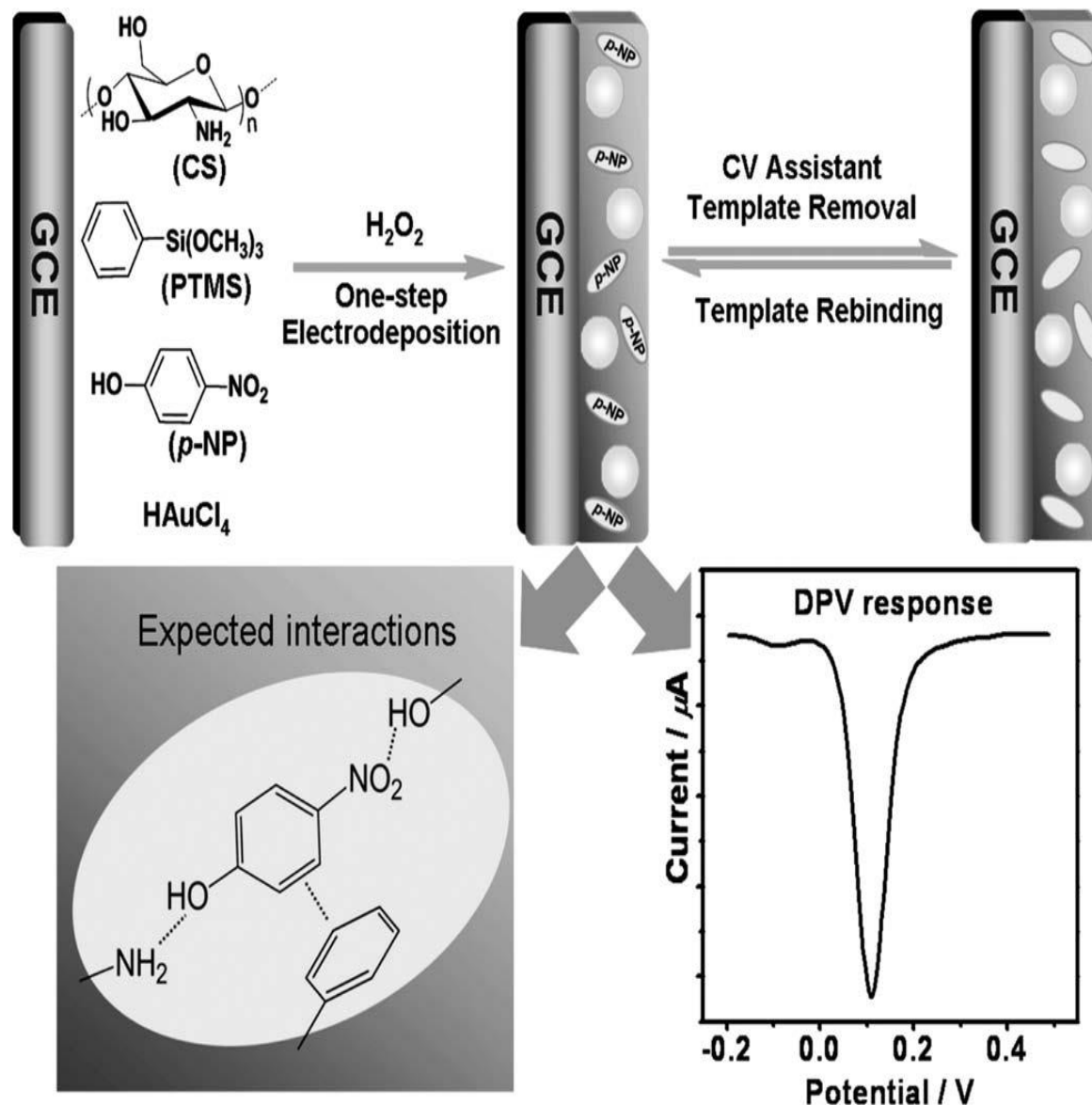
Laboratories and Analytical methods	LD2H GC/MS <sup>2</sup> (μg/L)	UME (μg/L)	C <sub>0</sub> (weighted) (μg/L)	
3 Samples	<b>PNP</b>			
	Estuarine	<b>114.5</b>	<b>117±6</b>	<b>108±6</b>
	Surface (dam)	<b>ND</b>	<b>106±6</b>	<b>104±6</b>
<b>MPT</b>	(GC/MS <sup>2</sup> )			
Estuarine	<b>93.8</b>	<b>96±3</b>	<b>120±6</b>	
Surface (dam)	<b>103.8</b>	<b>114±3</b>	<b>120±6</b>	

\*3 essays by samples

## 7- Improvement of the selectivity by the development of a MIP (= *molecular Imprinted Polymer* )

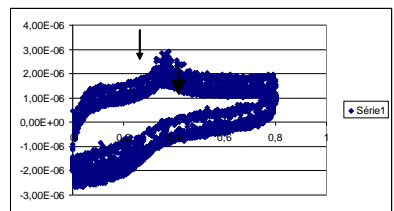
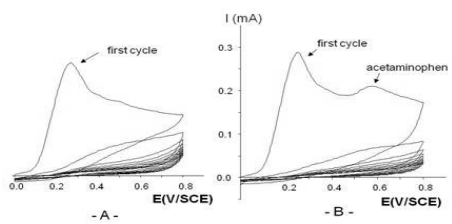
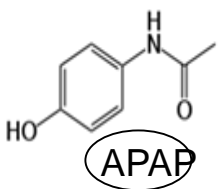
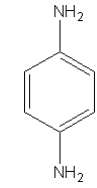
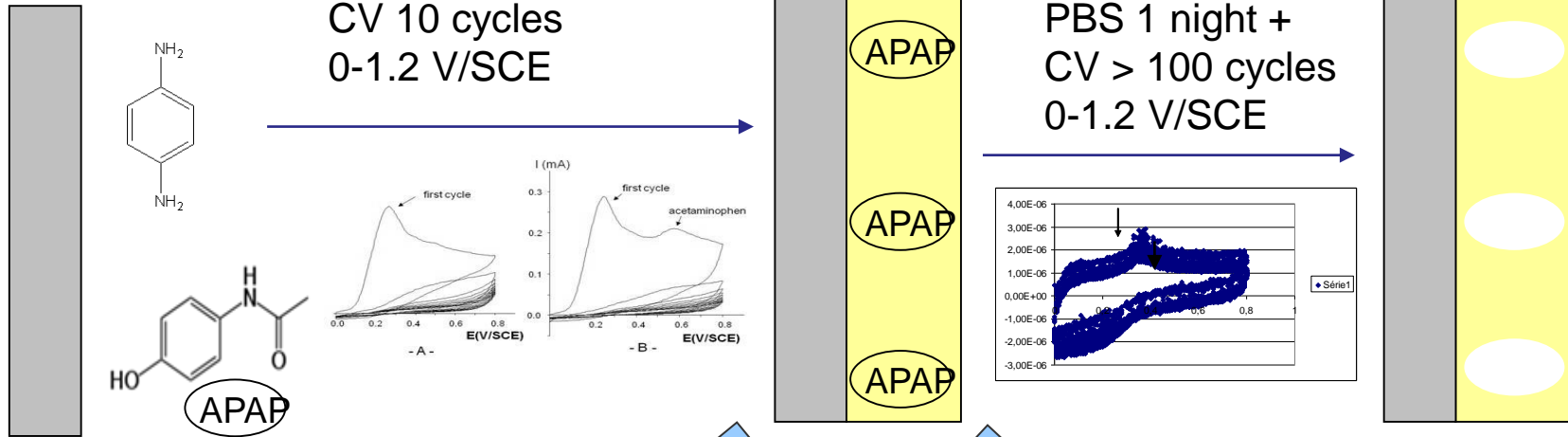


## MIP dedicated to PNP

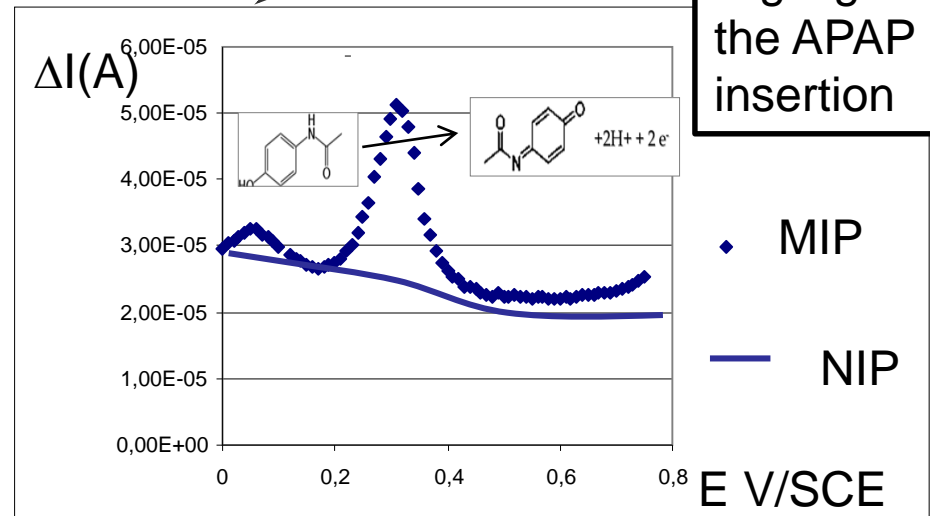
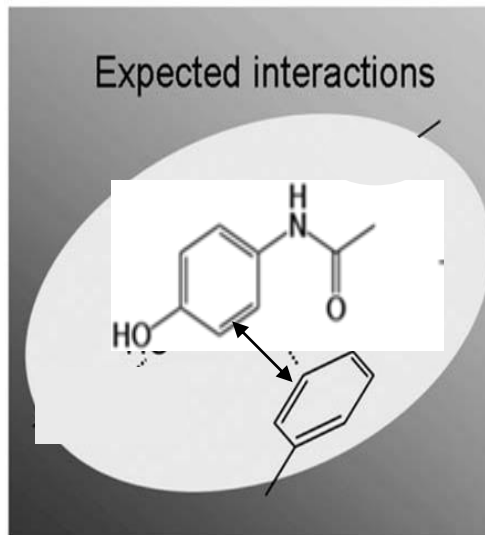


Schematic representation of the fabrication protocol of the p-NP imprinting and imprinted CS/PTMS/AuNPs/GCE for sensing p-NP. The one-step electrodeposition of the hybrid film on GCE was triggered by applying an optimal potential at -0.30 V vs. SCE for 300 s. The expected interactions in the recognition process involve hydrogen bonding and p-p stacking. The characteristic DPV response to p-NP locates at the **oxidation potential at +0.12 V vs. SCE** under the measurement conditions.

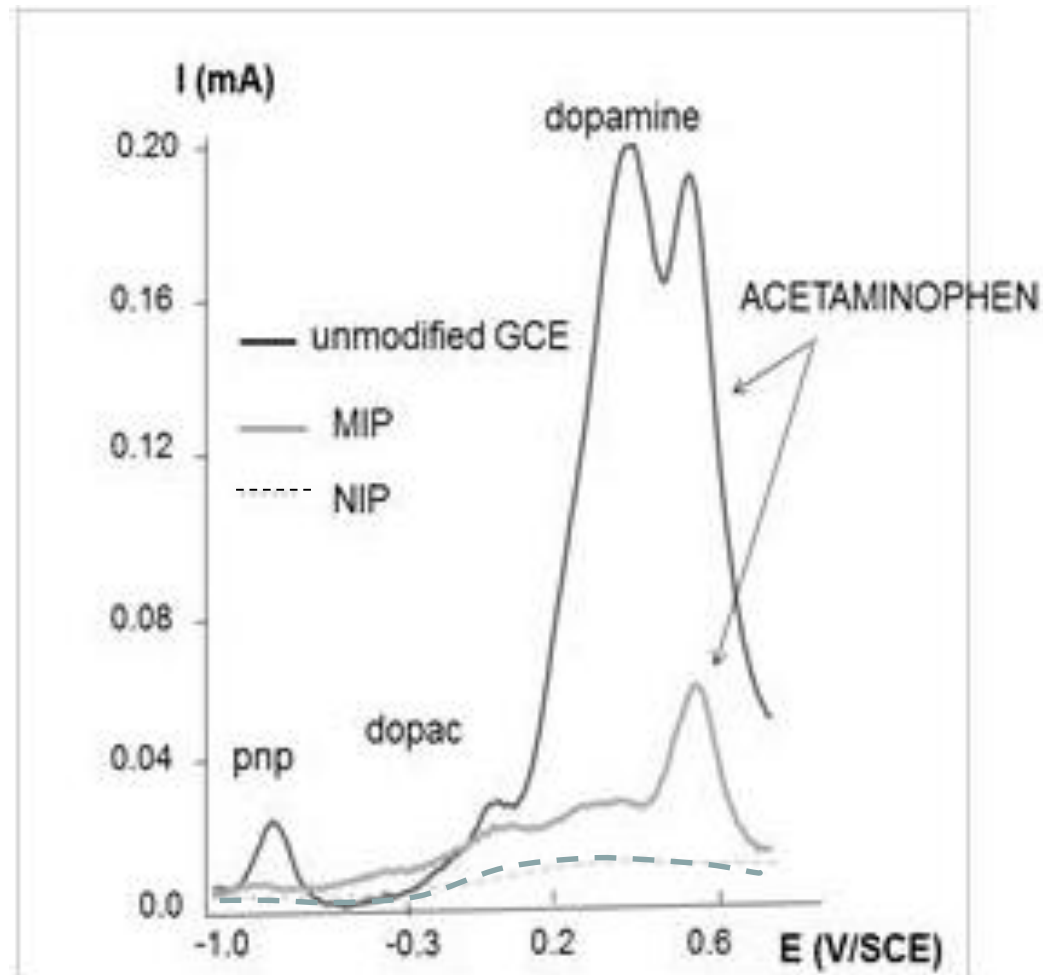
# MIP dedicated to acetaminophen



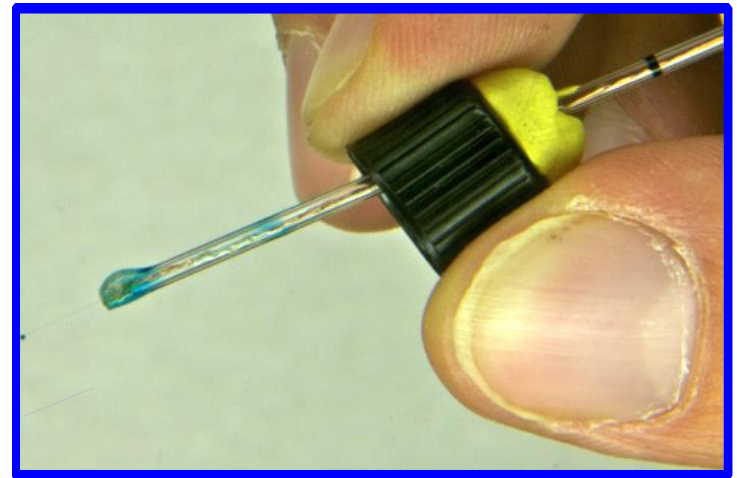
MM : 151.2 g/mol



# Interferences with ACETAMINOPHEN

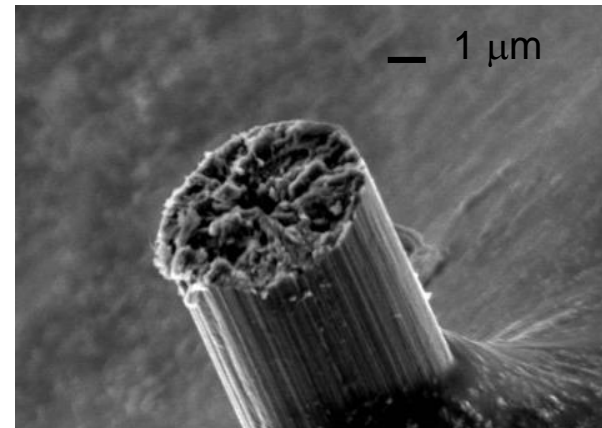


# 1<sup>er</sup> « *Homemade* » pesticide UME kit



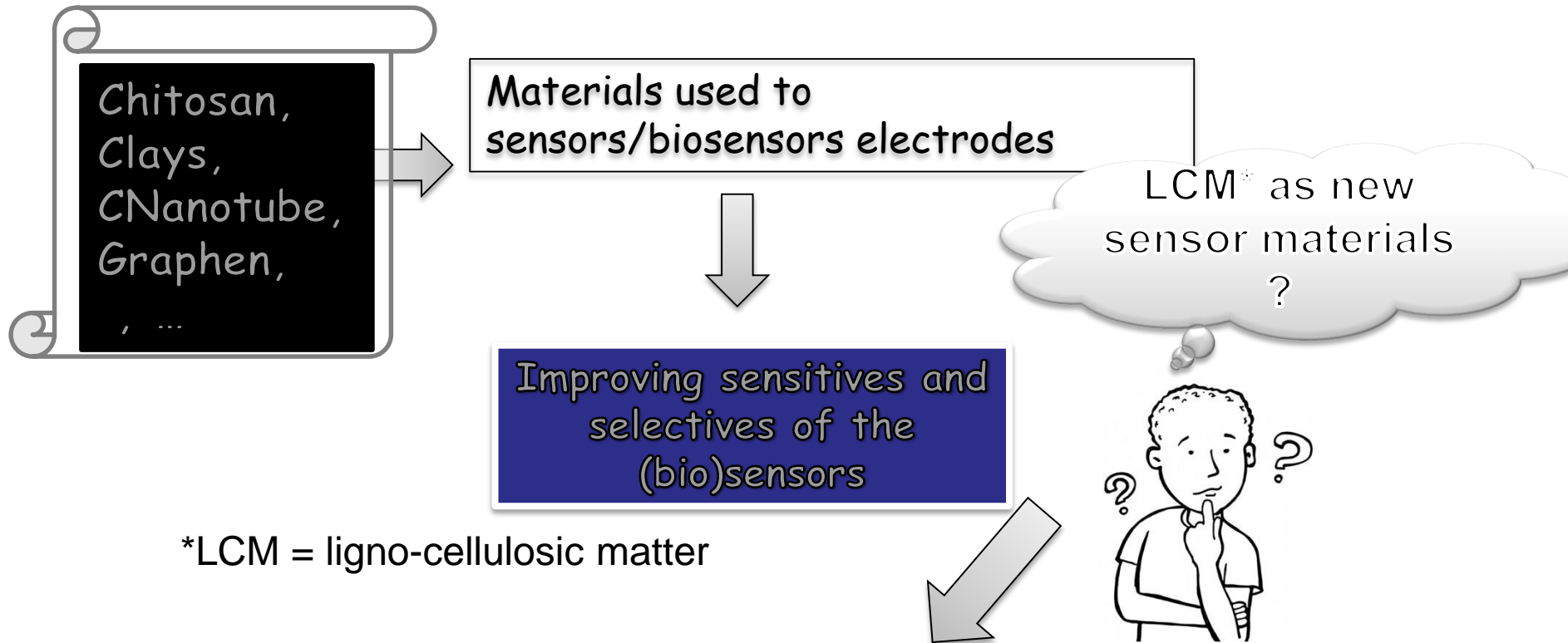
1<sup>er</sup> Kit de mesure des pesticides  
(Nanosenso™ 2010), [2]

*made in Angers*



[6] M. Pontié, « Techniques récentes en Analyse de l'Environnement et des Milieux Biologiques », Les Techniques de l'Ingénieur, IN 140, **2010**

# CPE acetaminophen sensitive sensor cellulose-based



\*LCM = ligno-cellulosic matter

Ecofriendly sensors, low cost analysis of drugs sold individually in the streets in Cameroon



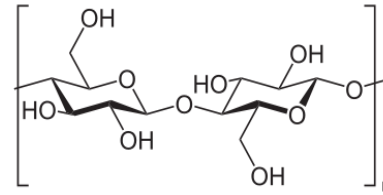
# LIGNO-CELLULOSIC MATERIALS



Coffee  
Husk (CH) from Cameroon

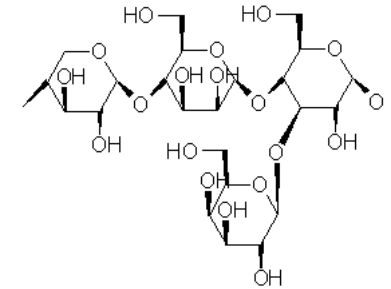


fraction size  
< 100  $\mu\text{m}$



cellulose

(40%)



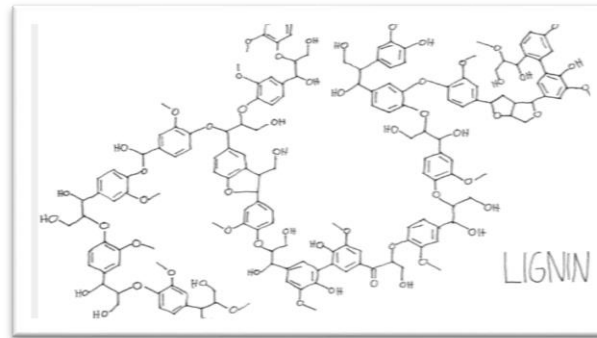
- Xylose -  $\beta(1,4)$  - Mannose -  $\beta(1,4)$  - Glucose -  
-  $\alpha(1,3)$  - Galactose

Hemi-cellulose ( $\approx 35\%$ )

**Extractibles matters** : polyphenols,  
phenols, pectins, sugars, esters of  
fatty acids, alcaloids, steroids...

(2 % to 8% of dry matter)

Minerals ( $\approx 2\%$ )



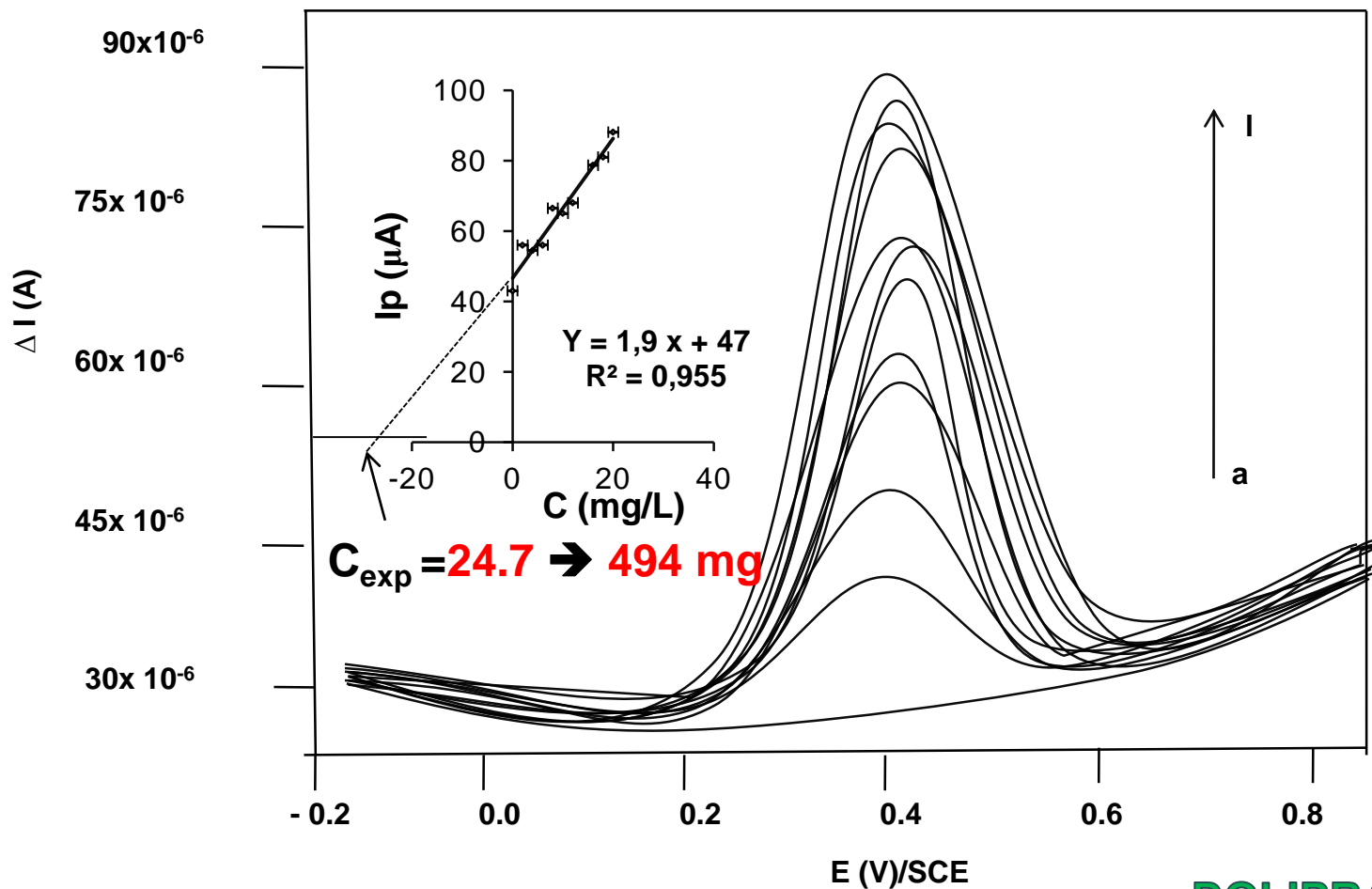
Lignin ( $\approx 17\%$ )

**55% cellulose**, 5% hemi-cellulose, 9% lignin, 31% others (not determined)





# CPE-CH validation with APAP commercialized tablets



**DOLIPRANE 500<sup>®</sup>**

# CPE-CH validation for DOLIPRANEs 500/1000 tablets

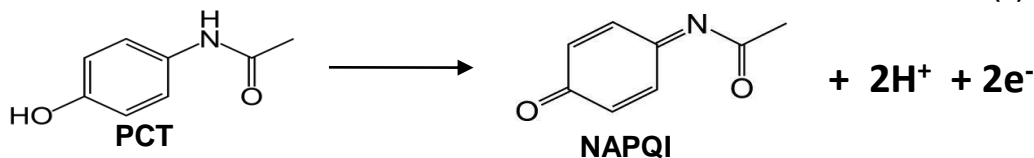
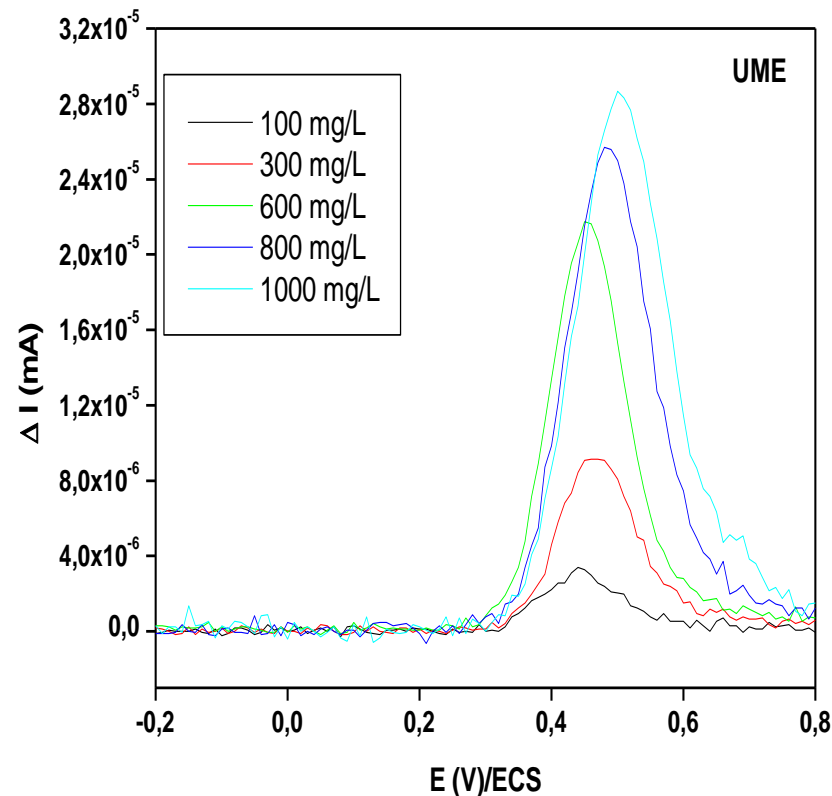
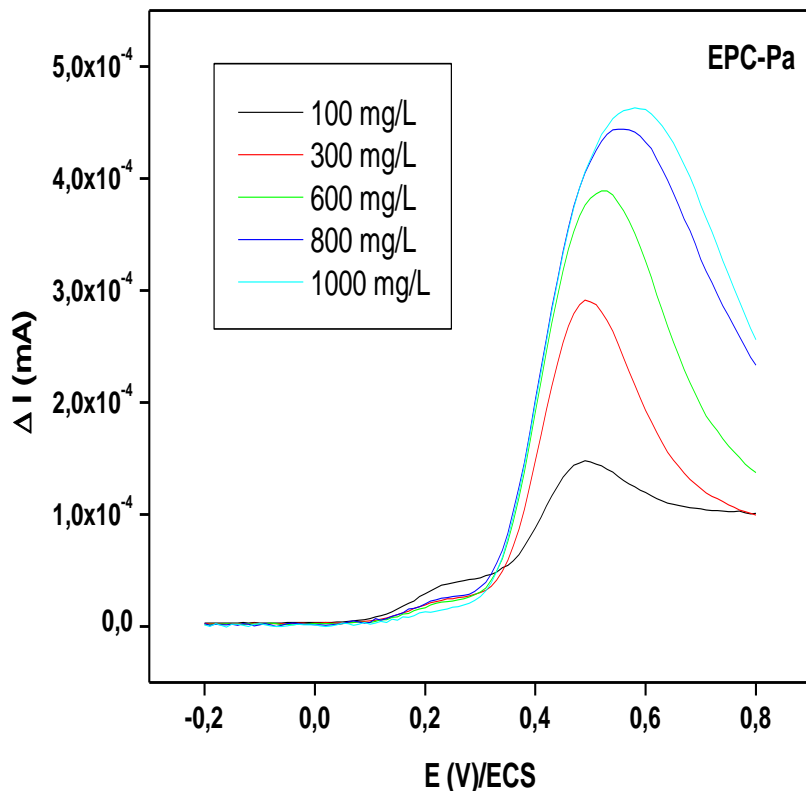
- Doliprane 500 ( Regulation : 500 ± **15 mg** )
- Doliprane 1000 ( Regulation : 1000 ± **30 mg** )

**3 tablets tested**

<b>Doliprane 500©</b> (weighted tablet # 600 mg)	slope( $\mu\text{A M}^{-1}$ )	1.9
	$R^2$	0.96
	$C_0$ (mg/L)	<b>24.7±0.5</b>
	Exp. APAP in tablets (mg)	<b>494 ± 9</b>
	Recovery (%)	97-101
<b>Doliprane 1000©</b> (weighted tablet # 1080 mg)	slope	2.1
	$R^2$	0.95
	$C_0$ (mg/L)	51.0±0.5
	Exp. APAP in tablets (mg)	<b>1020 ± 20</b>
	Recovery (%)	102-105

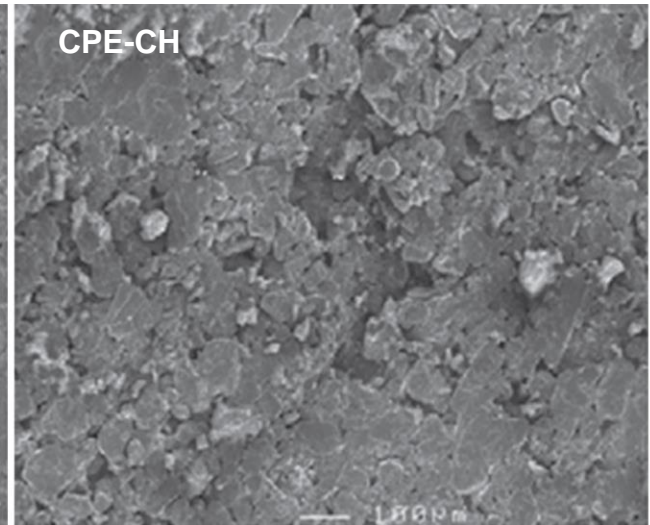
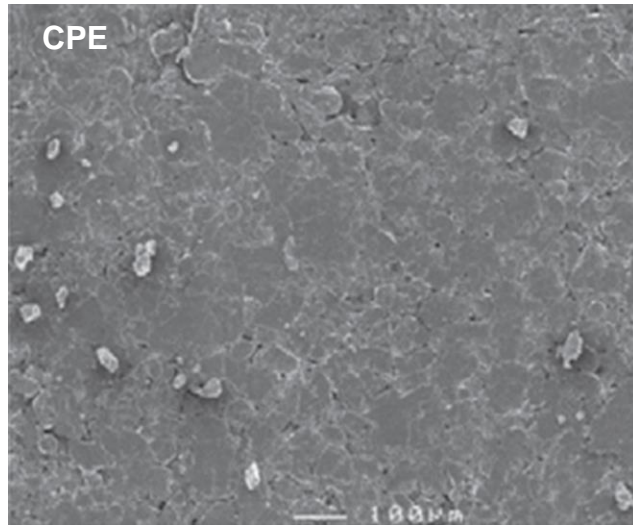
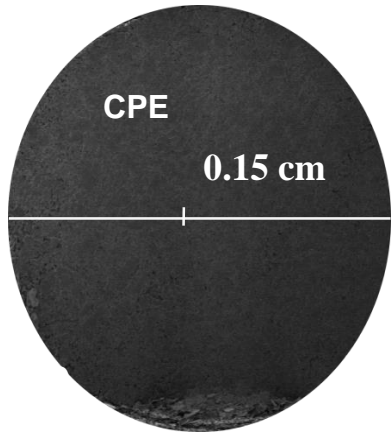
$$\text{Recovery} = (m_{\text{exp.}} / m_{\text{theory}}) * 100$$

# COMPARISON UME AND CPE vs APAP concentration



**Saturation** problem observed with CPE-CH from 600 mg/L, which is not the case of UME. **UME** is better adapted to "strong" concentration

# SEM analysis of CPEs modified and unmodified

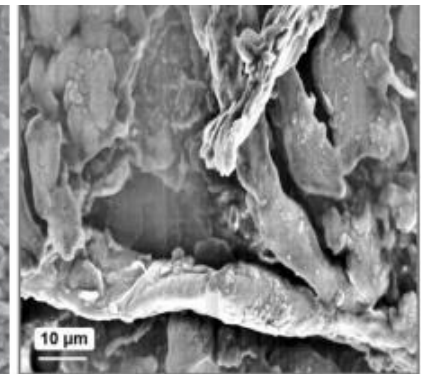
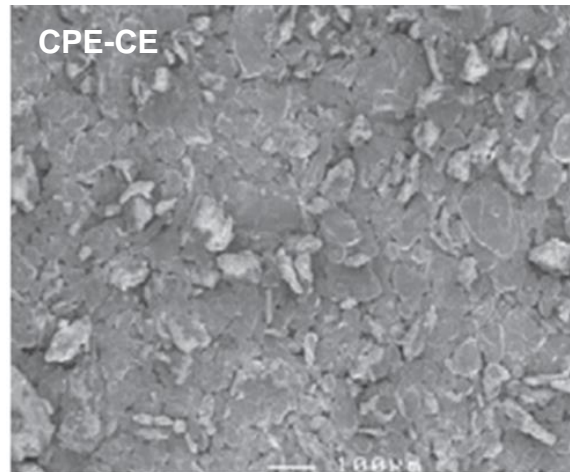


(magnification: x15)

Electrodes diameter = 3 mm

$$S_{\text{geom}} = \pi R^2 = 0.071 \text{ cm}^2$$

with  $R = 0.15 \text{ cm}$



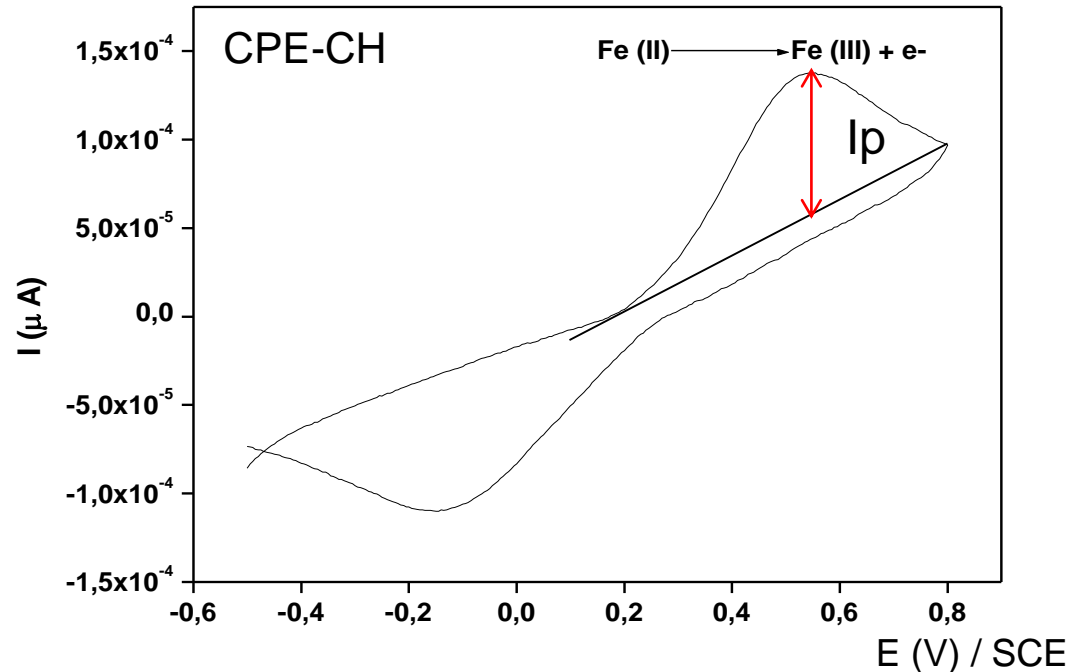
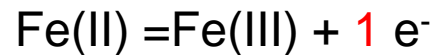
cellulose fibers length  
between 0.02-0.15 mm  
(Fluka)

*Coffee husk and cellulose particules change the morphology of CPE surface increasing the roughness*

# Determination of the real area with Fe(II) as a probe

Randles-Sevciks' law :

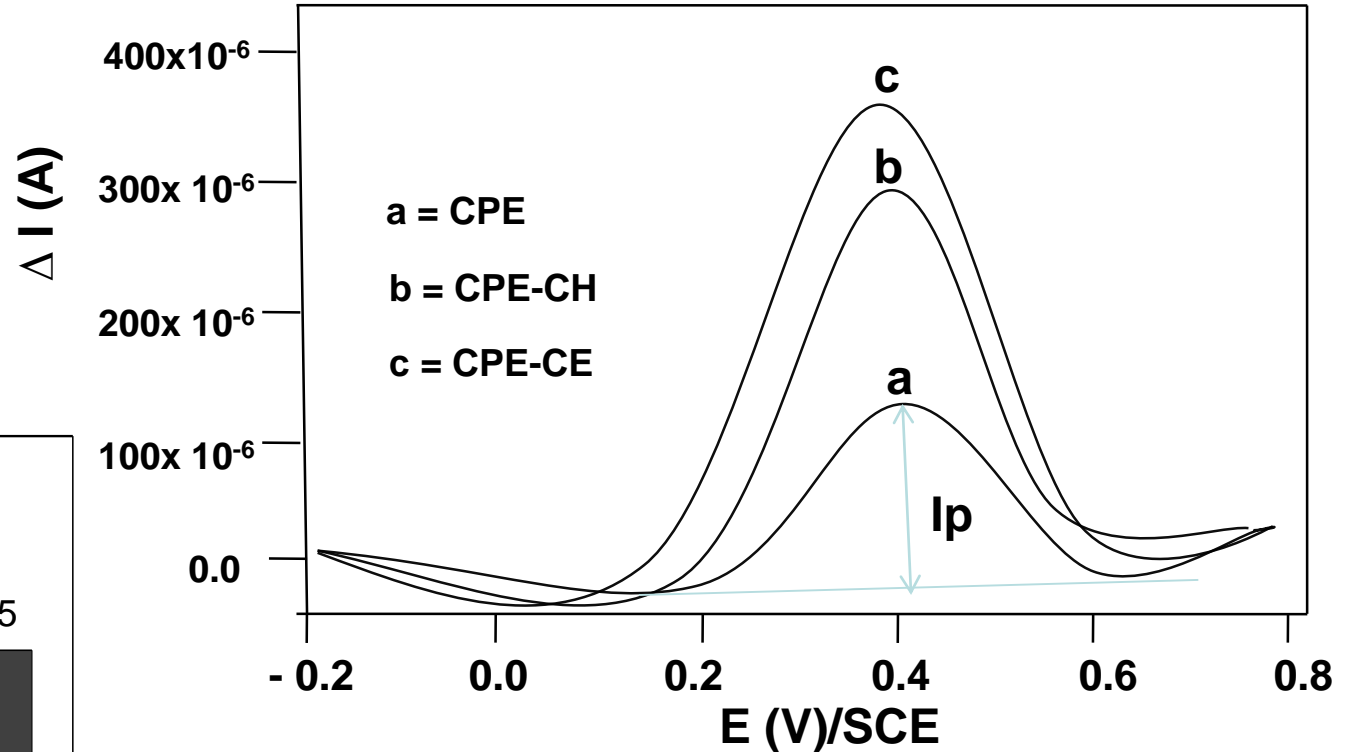
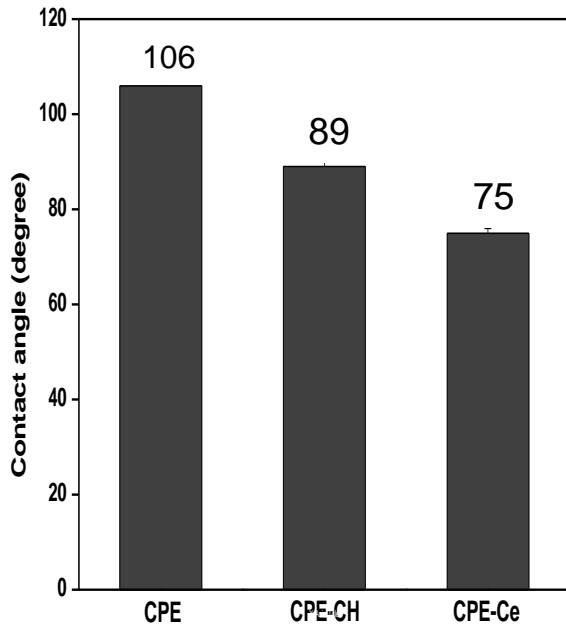
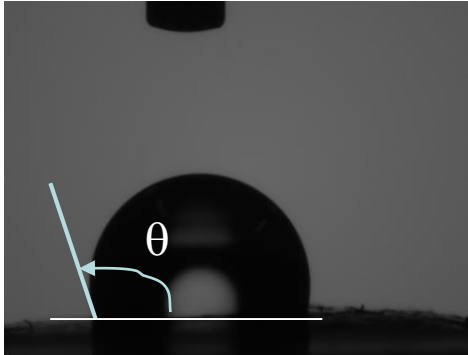
$$I_p = K n^{3/2} S D^{1/2} C V^{1/2}$$



Electrodes	Geometrical area	Real area (S)
CPE	0.071 cm <sup>2</sup>	0.082 cm <sup>2</sup>
CPE-CH	0.071 cm <sup>2</sup>	0.097 cm <sup>2</sup>
CPE-CE	0.071 cm <sup>2</sup>	0.141 cm <sup>2</sup>

$$S (\text{CPE-CE}) > S (\text{CPE-CH}) \gg S (\text{CPE})$$

# Correlation : sensitivity / hydrophilicity

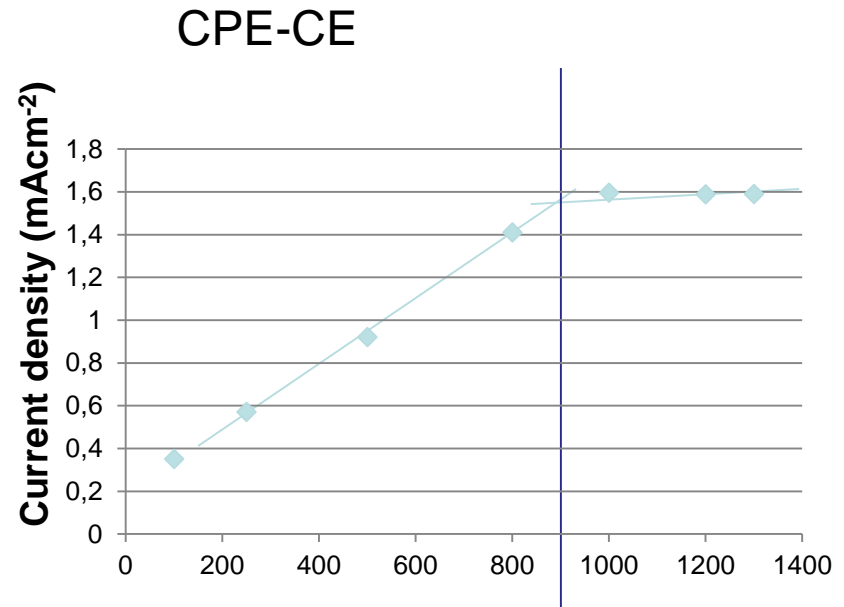
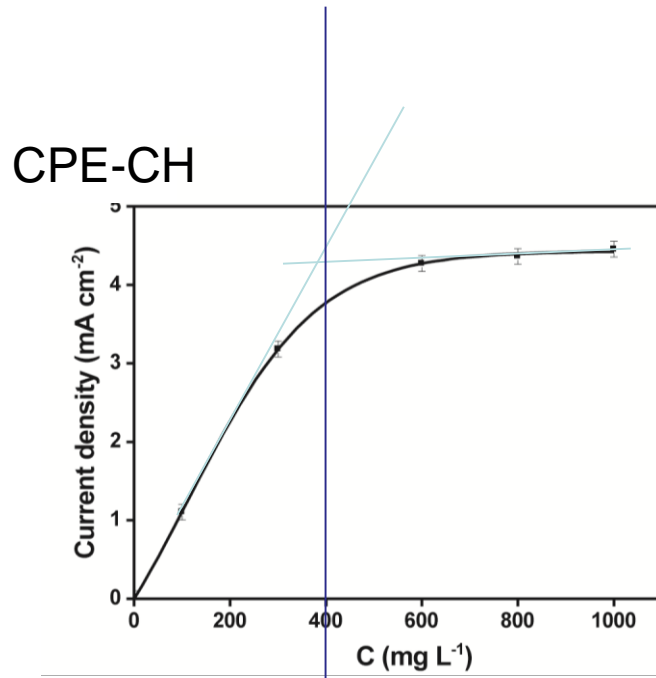


*Square wave voltamogram of APAP in PBS, pH=7.4, 0.1 M, 75 mg/L  
(pulse high : 90 mV; frequency : 400 Hz; scan Increment : 15 mV)*

Correlation between hydrophilicity increasement  
and peak intensity

**CPE-CE >> CPE-CH >> CPE**

# Area gain and limit of saturation for CPE-CH vs CPE-CE



Saturation : **400 mg/L**

**900 mg/L**

[8] Pontié, SF Mbokou, JP Bouchara, B Razafimandimby, S Egloff  
**Journal of Renewable Materials** 6 (3), (2018) 242-250

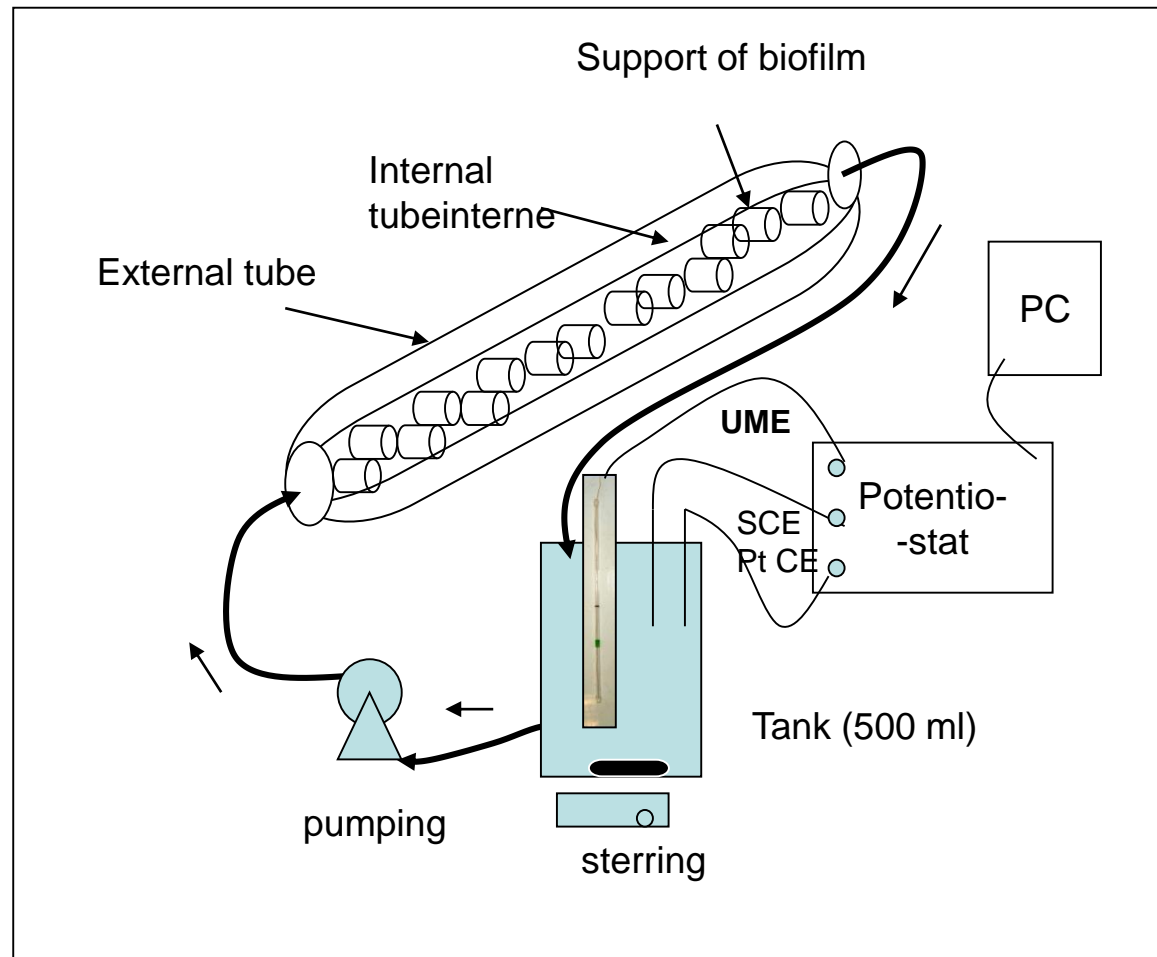
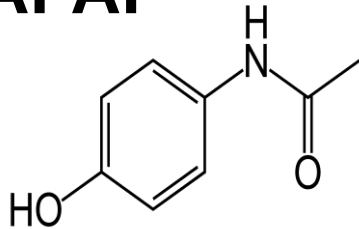


## **5/ CASE STUDIES 2 : Electrochemical analysis and WWT processes**

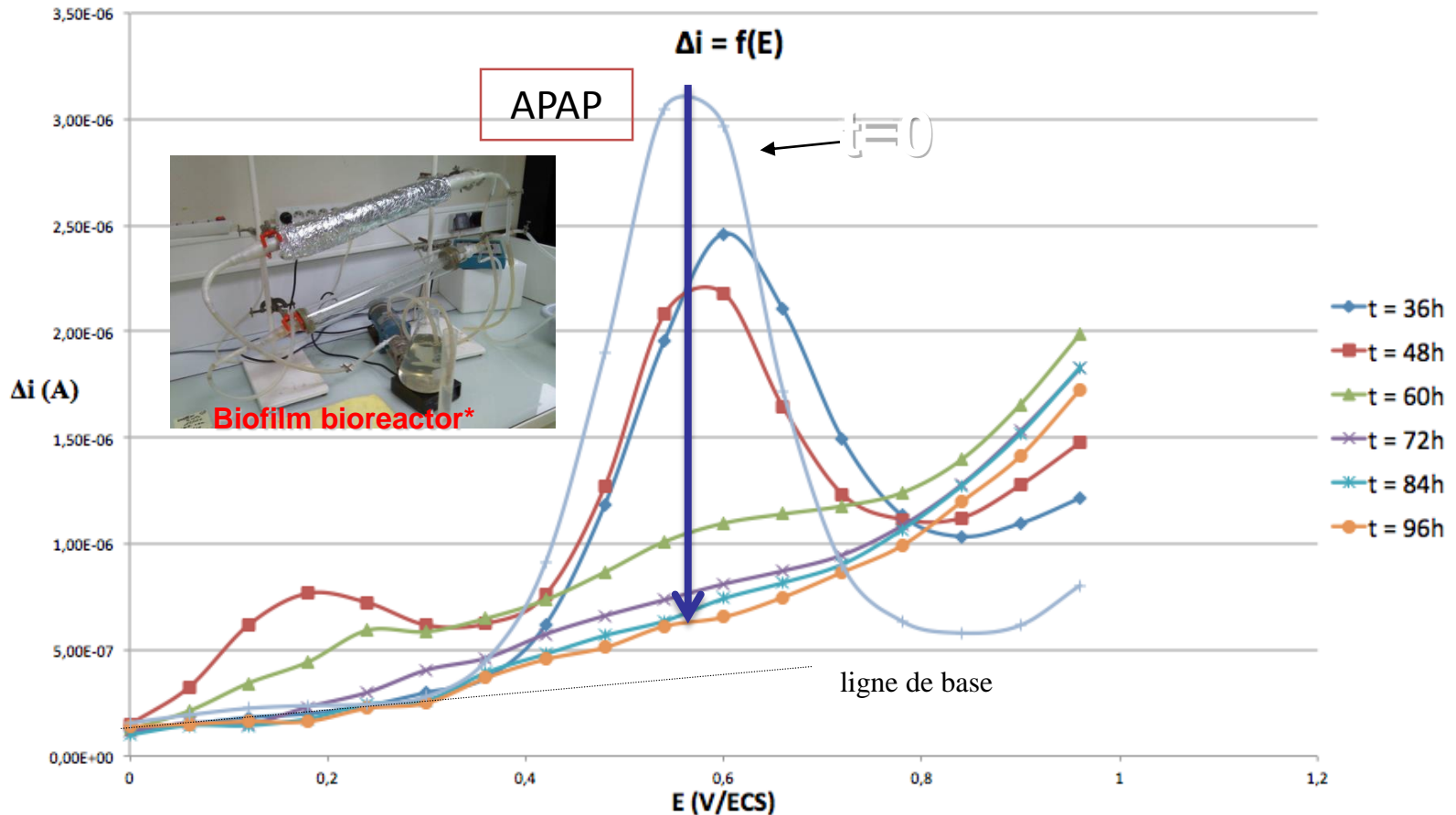
- **MEMBRANE BIOREACTOR FOR APAP BIODEGRADATION**
- STRATEGIES ANTIBIOFILMS ON UME vs CPE

# STUDY OF THE BIODEGRADATION OF APAP IN A BIOFILM BIOREACTOR

APAP



# BIODEGRADATION OF APAP

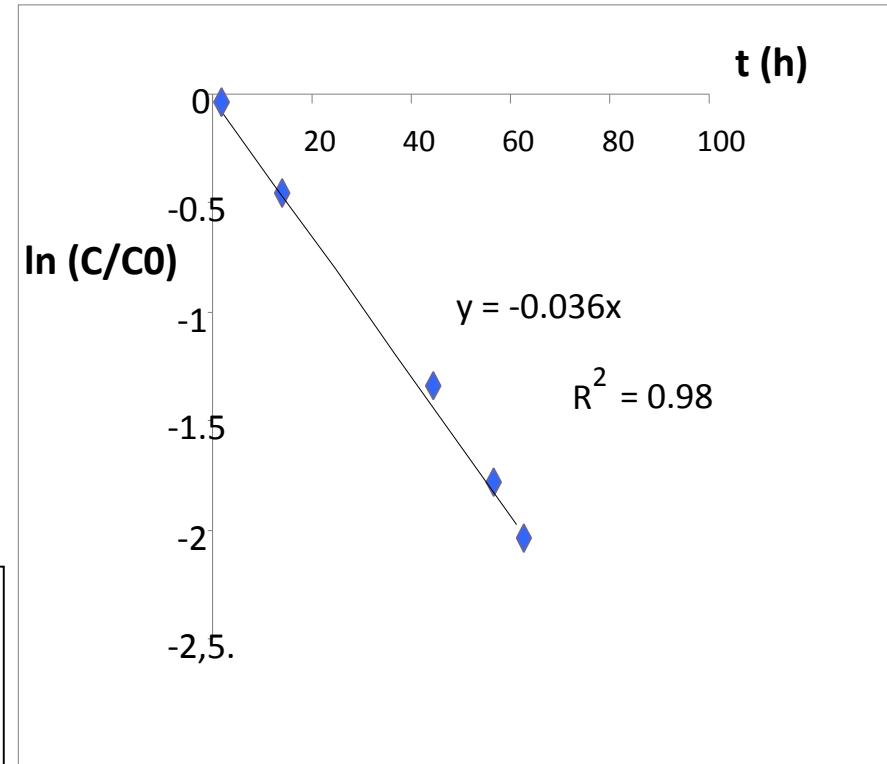
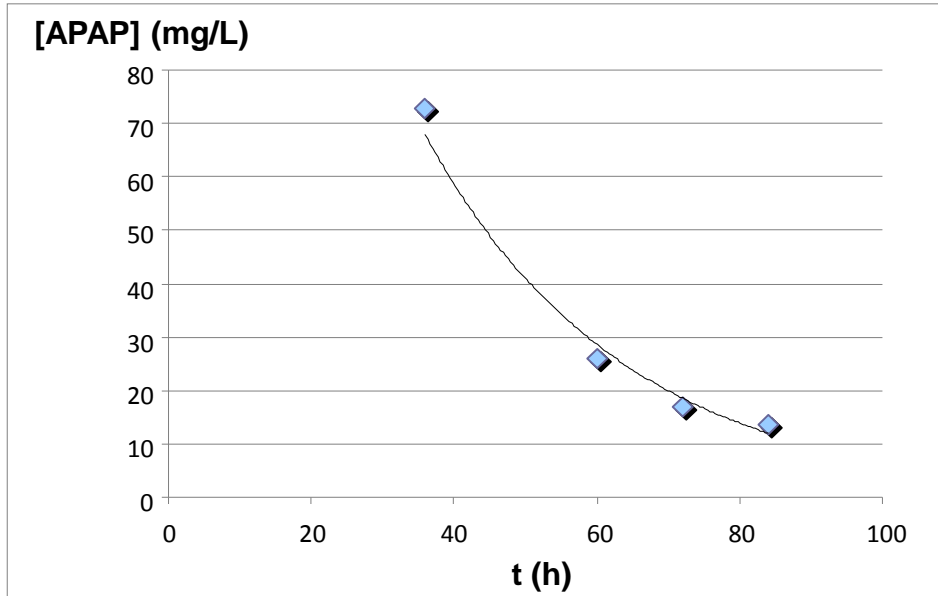


(\*biofilm of 8 months  
Thickness : 8.4 mm)

**$C_i = 100 \text{ mg/L}$**

**Conversion = 95% after 4 days**

# Kinetic parameters

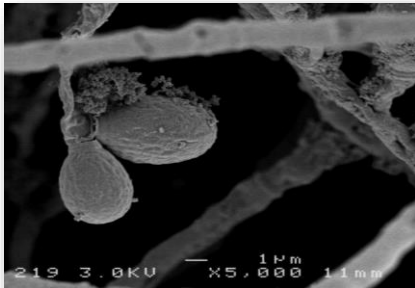
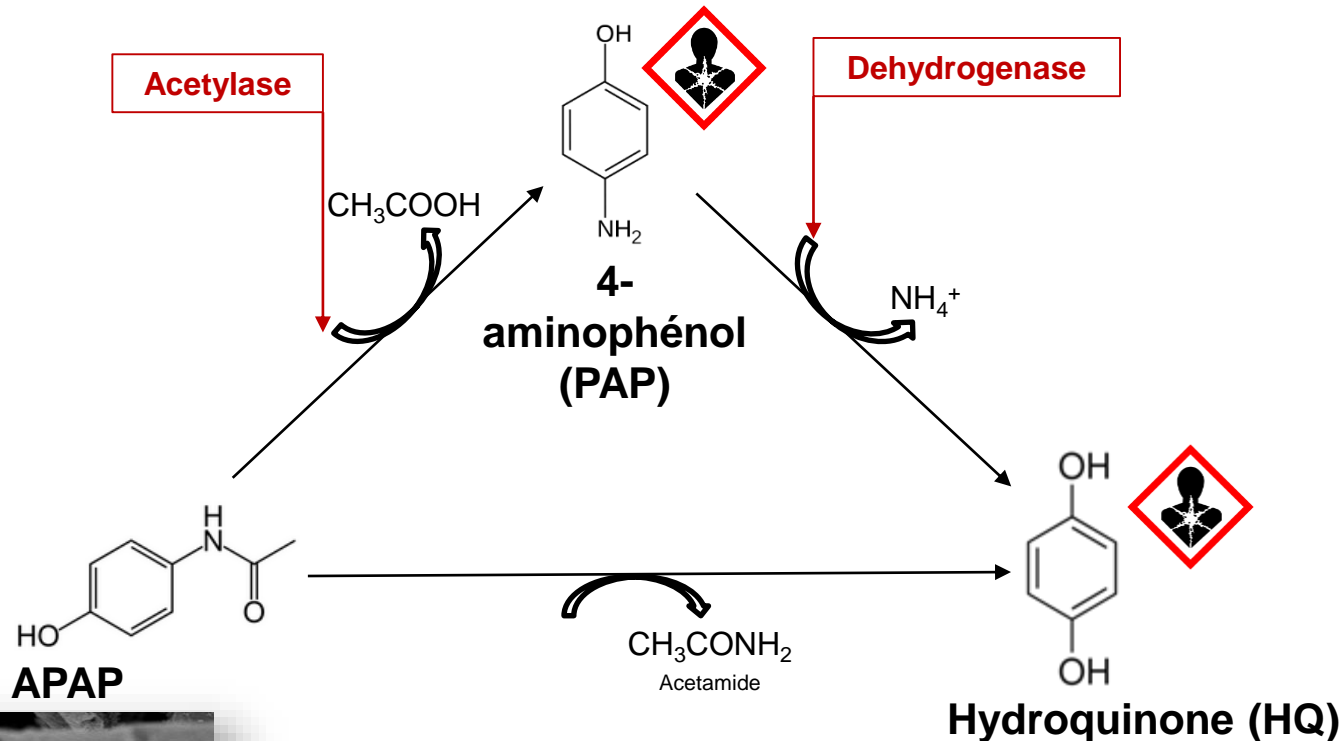


kinetic order 1 (T=20°C)

$$\ln C/C_0 = -0.036 \cdot t \quad R^2 = 0.96 \quad \text{and} \quad t_{1/2} = 19\text{h}$$

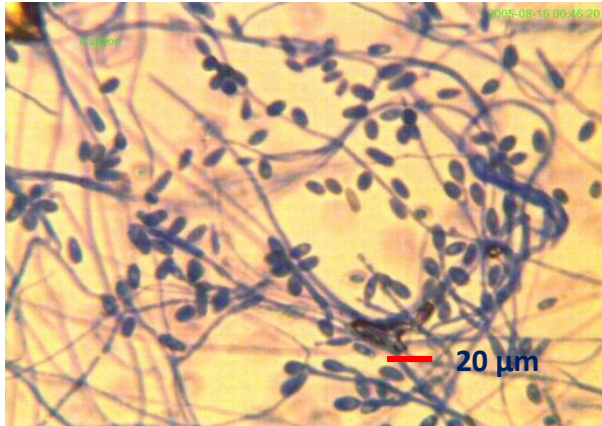
Conversion = 95% after 4 days

# APAP bacterial degradation by-products



**Another way** : find a microorganism usable for aromatic compounds biodegradation **with no formation of toxic by-products**

# *Scedosporium dehoogii*



*Scedosporium dehoogii* on YPD agar (14 days, 25°C).  
surface (left) and reverse (right) sides of the colony  
(strain n°110 350 905)

Optimal  
temperature 25°C but thermo-  
tolerant to 50°C

Saprophytic

Aero-anaeroby facultative

All the enzymatic equipment needed for hydrolysis  
of plant cell wall **lignin** and **polysaccharides**  
(cellulose, hemicellulose)

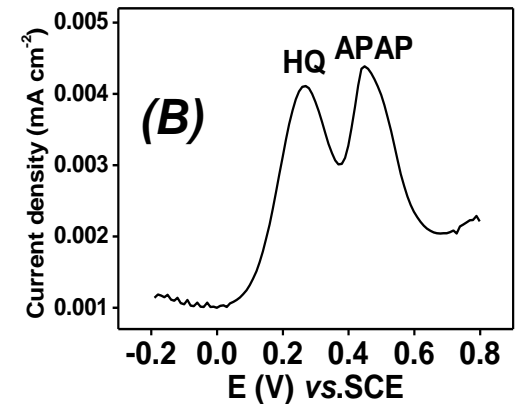
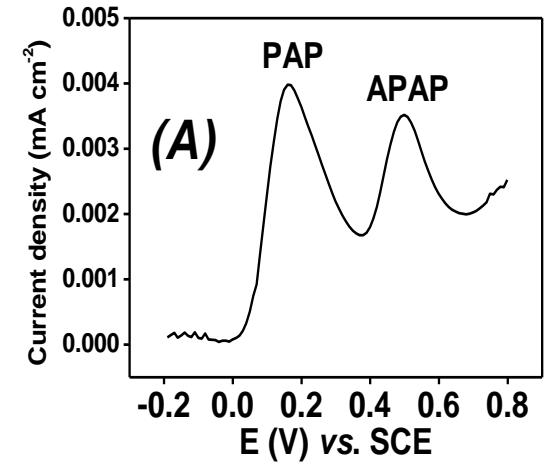
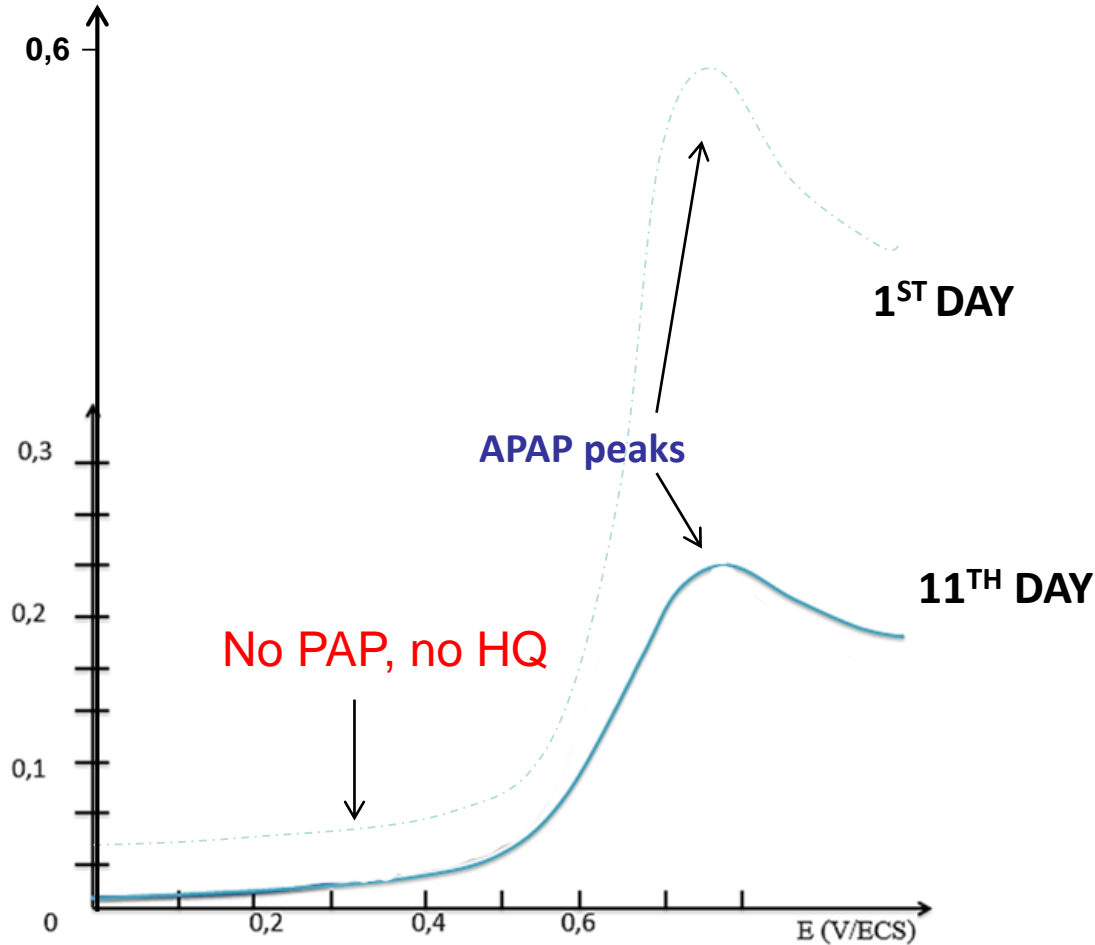
Encountered in highly polluted waters and soils  
(bioindicator of anthropogenic pollution)

Able to use **aromatic** or aliphatic **compounds**  
as sole carbon source

Several species pathogenic but not *S. dehoogii* !

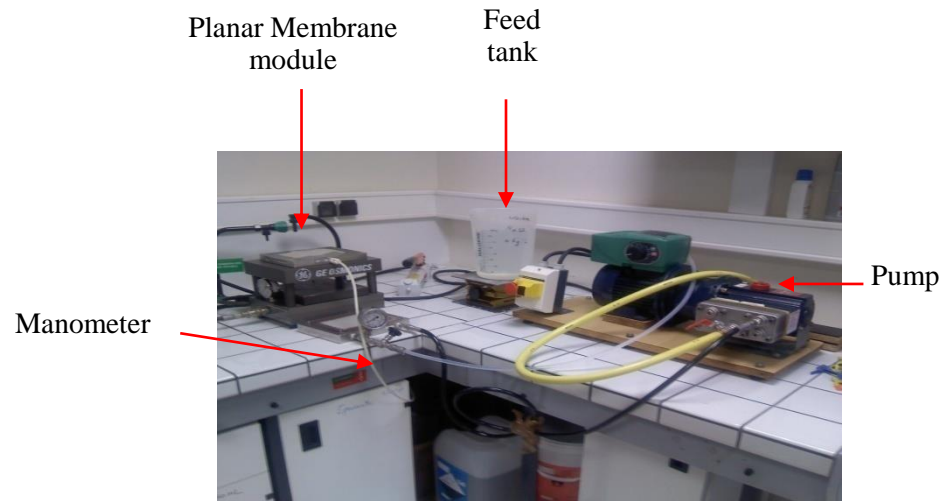
# APAP degradation by *Scedosporium dehoogii*

$\Delta I$  (mA)

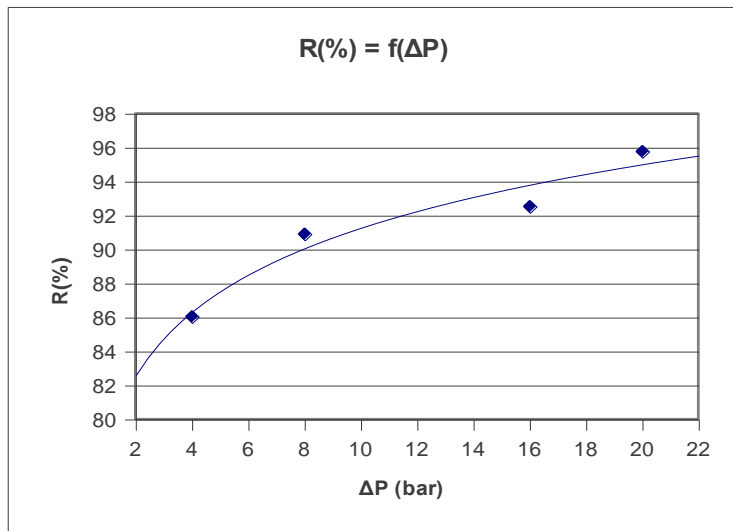


*S. dehoogii* is able to grow using APAP as source of carbon without toxic by-products like PAP or HQ

# NANOFILTRATION CAN ACHIEVE THE TREATMENT



NF lab scale device



Retention vs transmembrane pressure

At 20 bars, NF permeate gives **0.2 mg/L**  
(regulation effluents phenol index **< 0.3 mg/L**)

## CONCENTRATE SOLUTION MANAGED IN BIOREACTORS

[11] Nghiem L.D., Schafer A.I., Elimelech A.M. (2005).  
Environmental science & technology. Vol 39. 7698-7705.

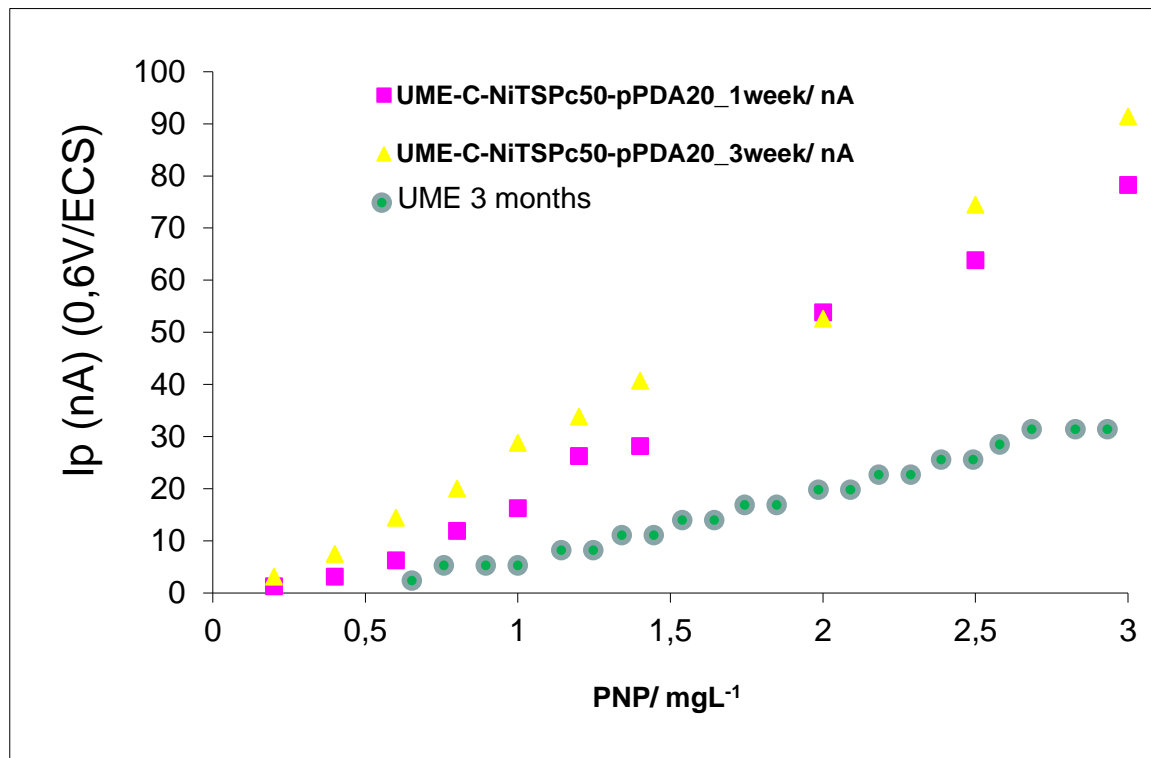


## **5/ CASE STUDIES 2 : Electrochemical analysis and WWT processes**

**- MEMBRANE BIOREACTOR FOR APAP BIODEGRADATION**

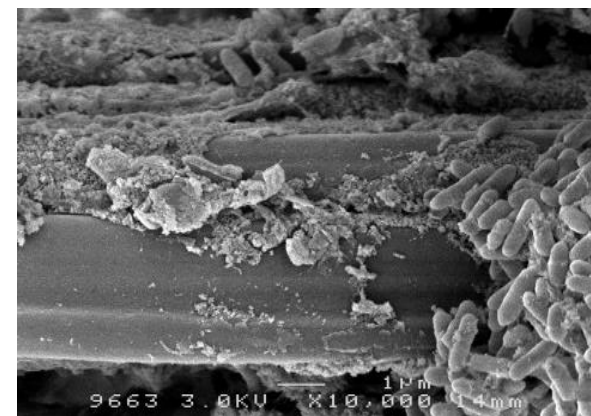
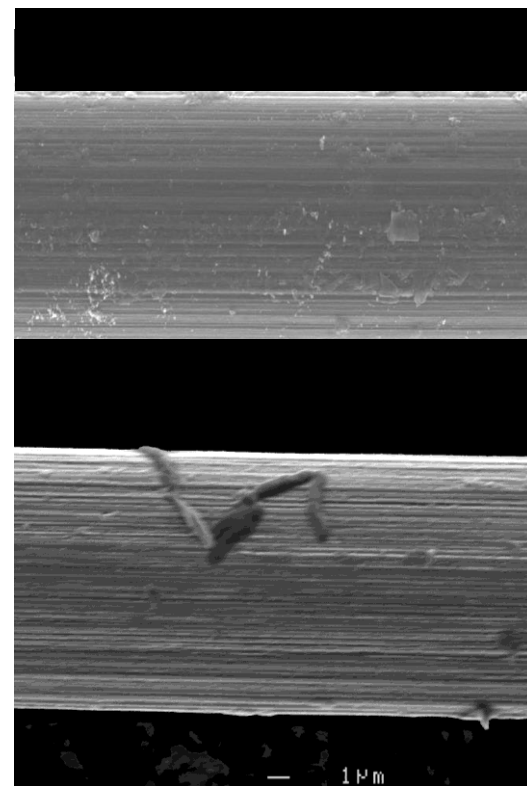
**- STRATEGIES ANTIBIOFILMS ON UME vs CPE**

# UME stocked 3 weeks in acetate buffer (pH=5,2) at ambient temperature and used during 3 months with real waters

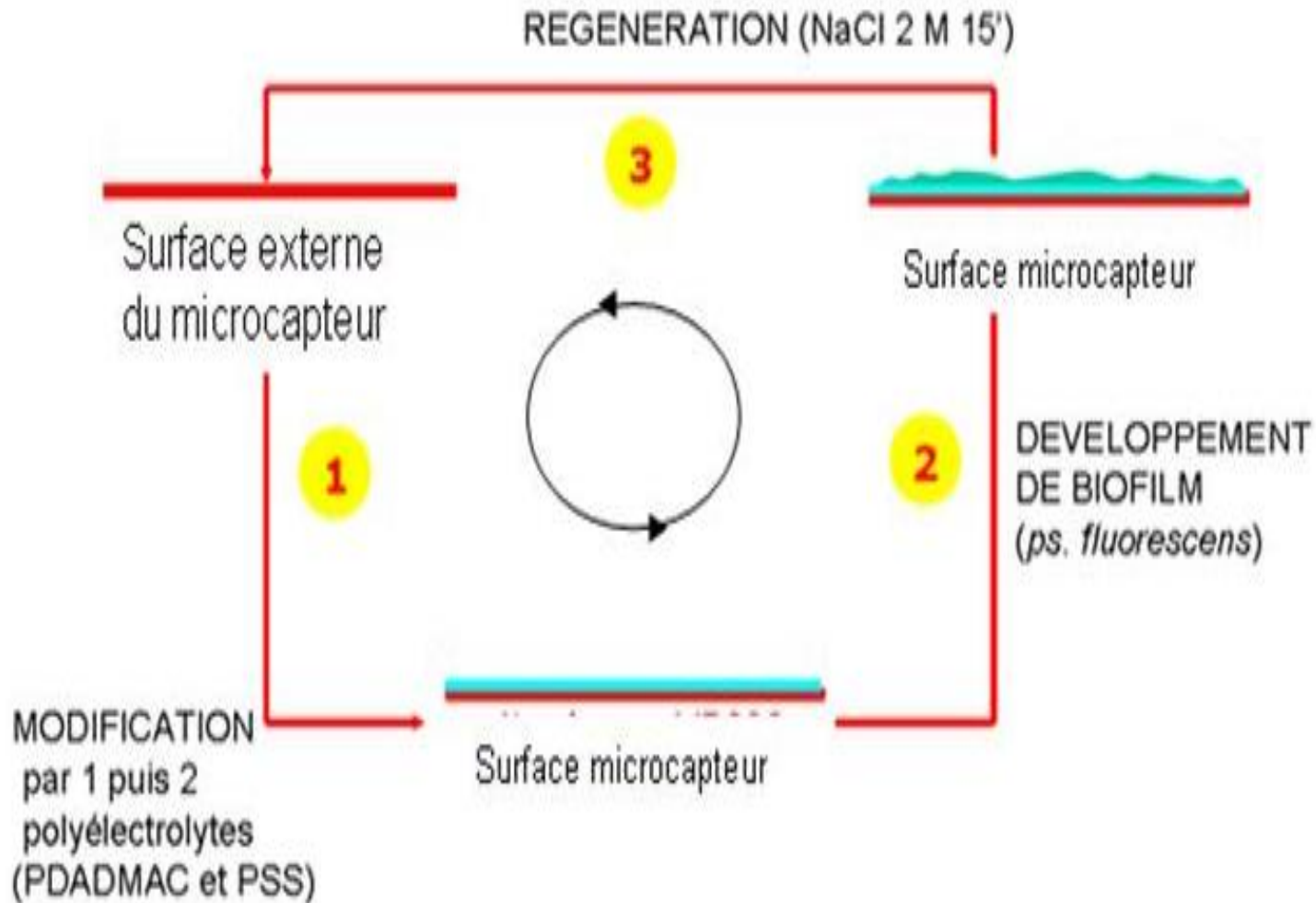


⇒ Only 5% lost of permeability + **microbial development start**

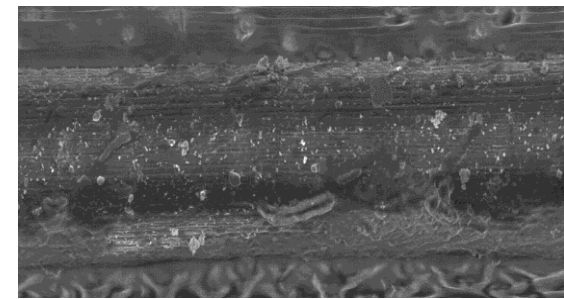
**After ACP biodegradation studies the bacterial colonisation was amplified**



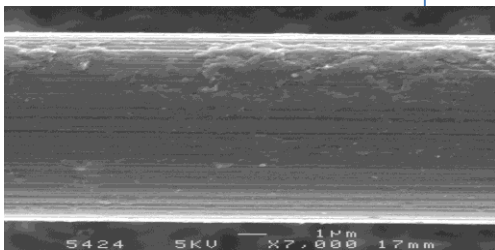
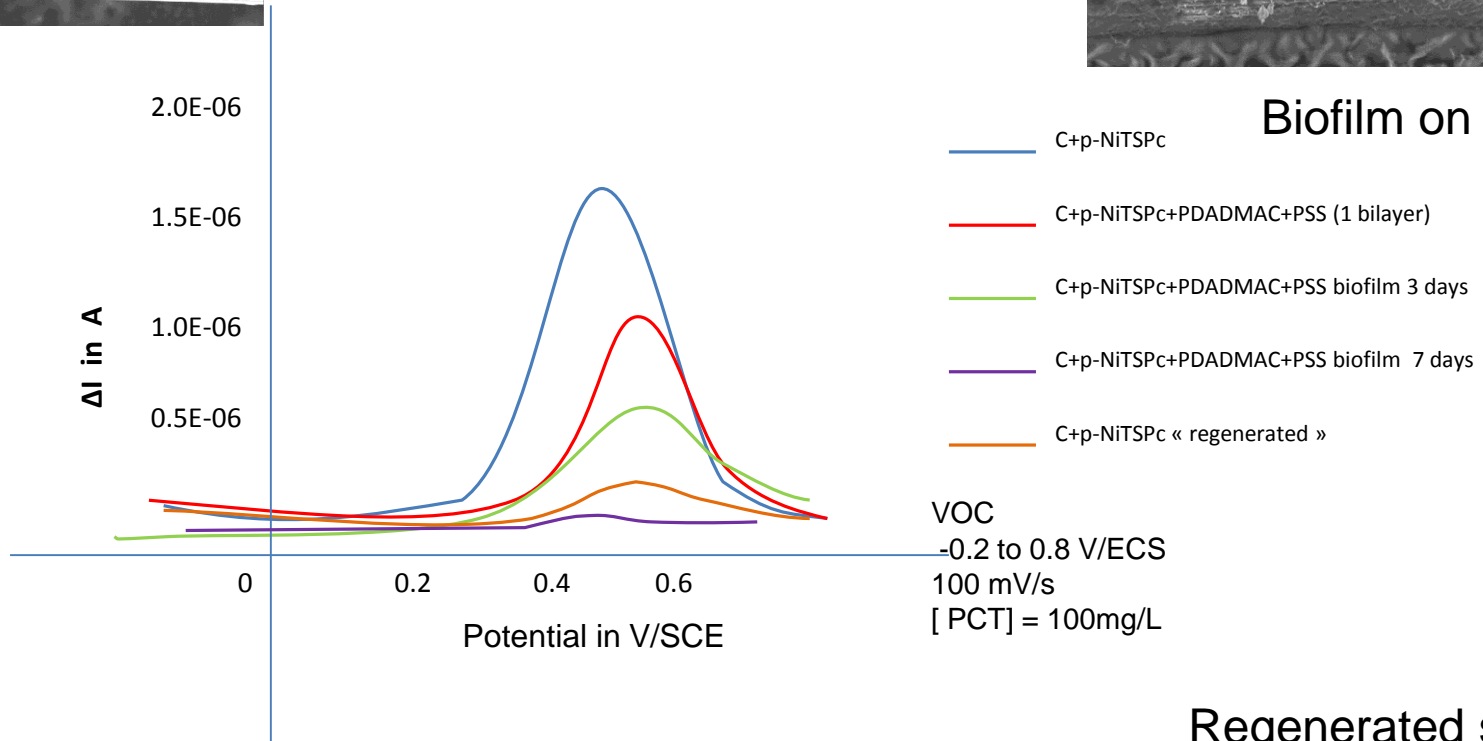
# ANTIBIOFOULING STRATEGY IN 3 STEPS



Pristine sensor



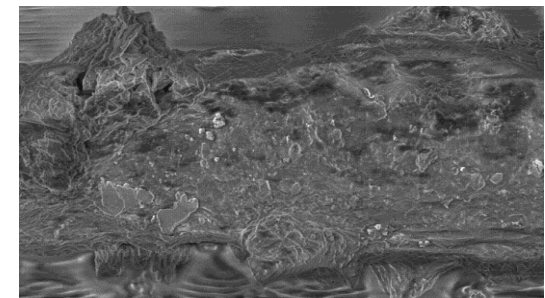
Biofilm on sensor



Sensor with 1 bilayer

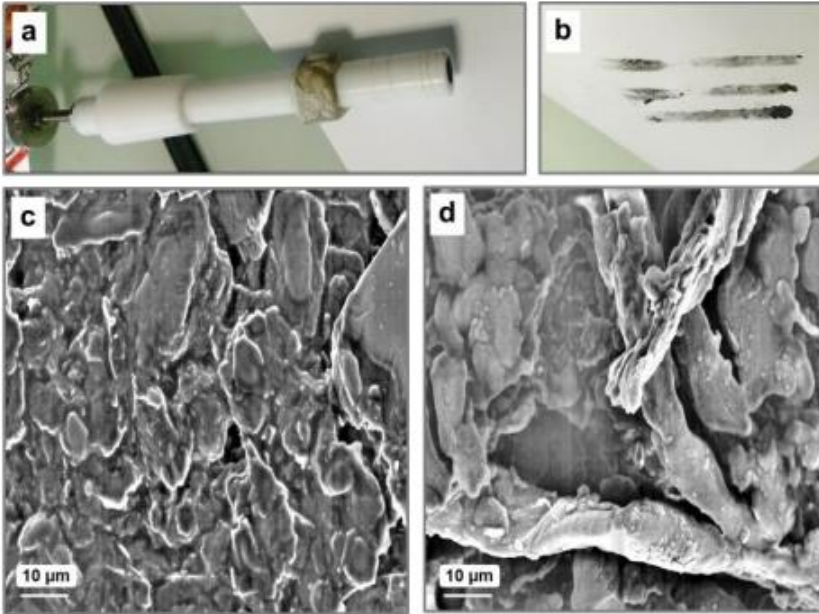
**VERY BAD REGENERATION**

**RESEARCH in development  
by testing multi-bilayers...**



Regenerated sensor

# An other antibiofouling strategy : Carbon paste electrode !



Sensitive  
« outlayer » **renewal**  
(3 lines of 3.5 cm) in 3  
seconds !

We have a « stock » of 30 electrodes



# Acknowledgements



ICMN team, Orléans

and PIERRE-YVES PONTALIER,  
LCA Toulouse



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WATER CELL, CAPACITES  
GEIHP Angers